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Orogeny forced terrestrial climate variation during the late Eocene–early Oligocene in Europe

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Table DR1. Background information on the sampled localities (1 page)

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Preparation of the samples for stable isotope analyses

Fig. DR1. Phosphate and structural carbonate oxygen isotopic compositions in biogenic apatite

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Fig. DR2. Detailed paleogeographic maps with tectonic units

References

TABLE DR1

| MAMMAL LEVEL | Map-Nr. | Locality | Country | Environment | Literature | Collections (specimens) |
|---------------------|----------------|----------------------------|----------------|-------------------------------------------------------|----------------------------------------------------------------------|--------------------------------|
| MP15+ | 1 | Csordakút | Hungary | marine - NP16 - Csolnok Claymarl Formation | Báldi-Beke, 1984; Ozsvárt, 1999; Kocsis, 2002 | MTM (1) |
| MP16 | 2 | Mormont-Station d'Eclépens | Switzerland | karst fissure fillings | Hooker & Weidmann, 2000 | MGL (8) |
| MP16 | 3 | St. Mamert du Gard | France | continental - stratified | Depéret & Carrière, 1901; Aguilar et al, 1997; | MTM (5) |
| MP17a | 4 | Euzet-les-Bains | France | continental - stratified | Aguilar et al, 1997; Paelobiology database - www.paleodb.org | MTM (5), NMB(5) |
| MP18 | 5 | La Débruge | France | mire or swamp - black sandy marl | Aguilar et al, 1997; Paelobiology database - www.paleodb.org | NMB (3) |
| MP18 | 6 | Gösgen-Canal | Switzerland | continental | Aguilar et al, 1997; Paelobiology database - www.paleodb.org | NMB (2) |
| MP18 | 7 | Râdaia (Andrásháza) | Romania | continental - Valea Nadășului Formation | Fărcaș and Codrea, 2008 | MAFI (1) |
| ?MP18-19 | 8 | Budapest - MTM collection | Hungary | ??? | Kretzoi, 1940 | MTM (1) |
| MP19 | 2 | Mormont-Eclépens C | Switzerland | karst fissure fillings | Hooker & Weidmann, 2000 | MGL (2), NMB (2) |
| MP19 | 9 | Liptingen 5 | Germany | karst fissure fillings | Scherzinger et al., 2005 | SMNS (4) |
| MP19 | 9 | Liptingen 13A | Germany | karst fissure fillings | Scherzinger et al., 2005 | SMNS (3) |
| MP19 | 10 | Neuhausen, Württemberg | Germany | karst fissure fillings | Scherzinger et al., 2005 | SMNS (3) |
| MP19 | 11 | Mähringen | Germany | karst fissure fillings | Scherzinger et al., 2005 | SMNS (4) |
| MP20 | 12 | Frohnstetten | Germany | karst fissure fillings | Scherzinger et al., 2005 | SMNS (6) |
| MP21- | 13 | Monteviale | Italy | continental - lignite deposit | Dal Piaz G.B., 1930, 1931; Tuchtan, 1953; Uhlig, 1999; Becker, 2009 | PGM (16) |
| MP21 | 14 | Ronzon | France | continental - lime mudstone - Phospherites du Quercy | Aguilar et al, 1997; Paelobiology database - www.paleodb.org | NMB (3) |
| MP21 | 15 | Möhren 19 | Germany | karst fissure fillings | Uhlig, 1999 | BSGP (11) |
| MP21-22 | 16 | Cluj | Romania | marine - NP22-23 - Mera Formation | Filipescu, 2001; Fărcaș and Codrea, 2008; Fărcaș, 2011 | MTM (1) |
| MP22 | 17 | Villebramar | France | continental - gravelly sand deposit | Brunet, 1970, 1979; Aguilar et al, 1997; Scherler, 2011 | NMB (14) |
| MP22 | 18 | Kleinblauen | Switzerland | marine - NP22 - Meeressand | Becker, 2009; Scherler, 2011 | NMB (1) |
| MP22 | 15 | Möhren13 | Germany | karst fissure fillings | Uhlig, 1999 | BSGP (15) |
| MP23+ | 19 | Budapest - Bohn clay pit | Hungary | marine - NP23 - top of the Tard Clay Formation | Kretzoi, 1940; Báldi-Beke, 1984; Báldi & Báldi-Beke, 1985 | MTM (1) |
| MP23-24 | 19 | Budapest - Ujlak clay pit | Hungary | marine - NP24 - Kiscell Clay Formation | Kretzoi, 1940; Báldi-Beke, 1984; Báldi & Báldi-Beke, 1985 | MTM (2) |
| MP23-24 | 16 | Cluj | Romania | continental - coal bearing depots - Dâncu Formation | Fărcaș and Codrea, 2008 | UBB (2) |
| MP23-24 | 20 | Aghires | Romania | continental - coal bearing depots - Dâncu Formation | Fărcaș and Codrea, 2008 | UBB (1) |
| MP23-25 | 21 | Csákberény | Hungary | continental - clay deposit - | Kretzoi, 1940 | MTM (2) |
| MP24- | 22 | Poillat | Switzerland | continental - Molasse alsacienne | Scherler, 2011; Becker et al., 2013 | MJSN (4) |
| MP24-25 | 23 | Tatabánya - Felsógalla | Hungary | continental - coal bearing depots - ?Csatka Formation | Kordos, 1978; Becker and Scherler (this study) | MTM (4) |
| MP25- | 24 | Bumbach | Switzerland | continental - Sub-alpine Molasse | Engesser and Mödden, 1997; Scherler, 2011; Scherler et al., in press | NMB (3) |

BSPG - Bayerische Staatssammlung für Paläontologie und Geologie (Bavarian State Collection for Palaeontology and Geology, Munich, Bavaria, Germany)

MAFI - Magyar Földtani Intezet (Collection of the Hungarian Geological Institute, Budapest, Hungary)

MGL - Musée géologique de Lausanne (Geological Museum, Lausanne, Switzerland)

MGP - Museo di Geologia e Paleontologia (Museum of Geology and Palaeontology Padua, Italy)

MJSN - Musée Jurassien des Sciences Naturelles (Jura Natural Science Museum, Porrentruy, Switzerland)

MTM - Magyar Természettudományi Muzeum (Hungarian Natural History Museum, Budapest, Hungary)

NMB - Naturhistorisches Museum Basel (Natural History Museum, Basel, Switzerland)

SMNS - Staatlichen Museum für Naturkunde Stuttgart (State Museum for Natural History, Stuttgart, Germany)

UBB - Fossil Collection of the University of Babes-Bolyai, Cluj, Romania

TABLE DR2

| Map-Nr. | Locality | Collection & Nr. | Taxon | Sampled material | Family | MAMMAL LEVEL | Age (Ma) | PHOSPHATE | | | | | | CARBONATE IN PHOSPHATE | | | | | | | | | | | |
|---------|------------------------------|------------------|-----------------|----------------------------------------------|--------------------|---------------------|----------|--------------------------------|-------|-----|--------|-----|---------|------------------------|-------|-----|-------|-----|---------|-------------------------------------|--------------------------------|---------------------------------|------|----------|-----|
| | | | | | | | | $\delta^{18}\text{O-PO4}$ SMOW | std.* | AV. | STD.** | N | 1.96xSE | $\delta^{13}\text{C}$ | std. | AV. | STD. | N | 1.96xSE | $\delta^{13}\text{C}_{\text{diet}}$ | $\delta^{18}\text{O-CO3}$ VPDB | $\delta^{18}\text{O-CO3}$ VSMOW | std. | CO3 wt.% | |
| 1 | Csordakút | HU | MTM-V.2002.12 | <i>Hyrachyus cf. stehlini</i> | left M1/2 | P - Hyrachyidae | MP15+ | 40.8 | 21.0 | 0.2 | 21.0 | 0.3 | 1 | 0.6 | -9.2 | 0.2 | -9.2 | 0.1 | 1 | 0.2 | -22.9 | -4.2 | 26.6 | 0.1 | 5.0 |
| 2 | Mormont - Station d'Éclépens | CH | MGL-2259 | <i>Palaeotherium curtum</i> | left m3 | P - Palaeotheriidae | MP16 | 39 | 21.7 | 0.1 | | | | | -7.6 | 0.0 | | | | | | -3.1 | 27.7 | 0.0 | 5.3 |
| 2 | Mormont - Station d'Éclépens | CH | MGL-2260 | <i>Palaeotherium curtum</i> | right m2 | P - Palaeotheriidae | MP16 | 39 | 19.2 | 0.1 | | | | | | | | | | | | | | | |
| 2 | Mormont - Éclépens A | CH | MGL-2391 | <i>Plagiolophus sp. 1</i> | left M3 | P - Pachynolophidae | MP16 | 39 | 22.3 | 0.2 | | | | | -7.0 | 0.1 | | | | | | -2.9 | 27.9 | 0.1 | 3.6 |
| 2 | Mormont | CH | MGL-2391 | <i>Plagiolophus stehlini</i> | right M3 | P - Palaeotheriidae | MP16 | 39 | 21.0 | 0.2 | | | | | -7.0 | 0.0 | | | | | | -4.3 | 26.5 | 0.1 | 5.4 |
| 2 | Mormont - Station d'Éclépens | CH | MGL-1620 | <i>Lophiodon lautricense</i> | right P3 | P - Lophiodontidae | MP16 | 39 | 21.3 | 0.0 | | | | | -8.2 | 0.1 | | | | | | -3.7 | 27.1 | 0.1 | 4.9 |
| 2 | Mormont - Station d'Éclépens | CH | MGL-1623 | <i>Lophiodon lautricense</i> | left m2 | P - Lophiodontidae | MP16 | 39 | 21.5 | 0.2 | | | | | -8.7 | 0.1 | | | | | | -4.5 | 26.3 | 0.1 | 1.9 |
| 2 | Mormont - Station d'Éclépens | CH | MGL-1627 | <i>Lophiodon tapiroides</i> | right p2 | P - Lophiodontidae | MP16 | 39 | 21.4 | 0.3 | | | | | -8.5 | 0.1 | | | | | | -5.2 | 25.6 | 0.1 | 2.2 |
| 2 | Mormont - Station d'Éclépens | CH | MGL-1628 | <i>Lophiodon tapiroides</i> | right M2 (M1?) | P - Lophiodontidae | MP16 | 39 | 21.2 | 0.2 | 21.2 | 0.9 | 8 | 0.6 | -8.7 | 0.1 | -8.0 | 0.7 | 7 | 0.5 | -21.8 | -4.8 | 25.9 | 0.1 | 1.8 |
| 3 | St. Mamert du Gard | FR | MTM-V.60.152 | <i>Lophiodon franconica</i> | left M3 | P - Lophiodontidae | MP16 | 38.85 | 21.6 | 0.1 | | | | | | | | | | | | | | | |
| 3 | St. Mamert du Gard | FR | MTM-V.60.153 | <i>Lophiodon franconica</i> | right M2/3 | P - Lophiodontidae | MP16 | 38.85 | 21.1 | 0.0 | | | | | -9.2 | 0.1 | | | | | | -6.2 | 24.5 | 0.2 | 2.4 |
| 3 | St. Mamert du Gard | FR | MTM-V.60.151 | <i>Lophiodon franconica</i> | tooth frg. m1(m2?) | P - Lophiodontidae | MP16 | 38.85 | 21.8 | 0.1 | | | | | | | | | | | | | | | |
| 3 | St. Mamert du Gard | FR | MTM-V.60.187 | <i>Lophiodon franconica</i> | left m1? | P - Lophiodontidae | MP16 | 38.85 | 21.6 | 0.2 | 21.5 | 0.3 | 4 | 0.3 | | | -9.2 | 0.1 | 1 | 0.2 | -23.0 | | | | |
| 4 | Euzet les Bains | FR | NMB-no number | <i>Plagiolophus sp. (annectens ?)</i> | left M2 | P - Pachynolophidae | MP17a | 37.5 | 20.3 | 0.1 | | | | | -7.0 | 0.0 | | | | | | -6.6 | 24.1 | 0.0 | 5.9 |
| 4 | Euzet les Bains | FR | NMB-no number | <i>Plagiolophus sp. (annectens ?)</i> | left M3 | P - Pachynolophidae | MP17a | 37.5 | 22.6 | 0.2 | | | | | -7.3 | 0.0 | | | | | | -4.7 | 26.1 | 0.1 | 6.5 |
| 4 | Euzet les Bains | FR | NMB-no number | <i>Plagiolophus sp. (annectens ?)</i> | M | P - Pachynolophidae | MP17a | 37.5 | 19.9 | 0.2 | | | | | -7.8 | 0.1 | | | | | | -3.9 | 26.8 | 0.1 | 3.9 |
| 4 | Euzet les Bains | FR | NMB-no number | <i>Plagiolophus sp. (annectens ?)</i> | M | P - Pachynolophidae | MP17a | 37.5 | 22.0 | 0.1 | | | | | -9.1 | 0.1 | | | | | | -3.3 | 27.5 | 0.1 | 3.5 |
| 4 | Euzet les Bains | FR | NMB-no number | <i>Plagiolophus sp. (annectens ?)</i> | m | P - Pachynolophidae | MP17a | 37.5 | 19.7 | 0.1 | | | | | -8.7 | 0.1 | | | | | | -6.3 | 24.4 | 0.0 | 6.3 |
| 4 | Euzet les Bains | FR | MTM-V.60.84 | <i>Lophiotherium cervulum</i> | left m2 | P - Lophiodontidae | MP17a | 37.5 | 19.6 | 0.2 | | | | | -7.8 | 0.1 | | | | | | -4.4 | 26.3 | 0.1 | 4.2 |
| 4 | Euzet les Bains | FR | MTM-V.60.1317 | <i>Palaeotherium crassum</i> | left P4 | P - Palaeotheriidae | MP17a | 37.5 | 20.5 | 0.3 | | | | | | | | | | | | | | | |
| 4 | Euzet les Bains | FR | MTM-V.60.82 | <i>Anchilophus dumasi</i> | right M3 | P - Palaeotheriidae | MP17a | 37.5 | 21.4 | 0.1 | | | | | | | | | | | | | | | |
| 4 | Euzet les Bains | FR | MTM-V.60.1319 | <i>Paloplotherium annectens</i> | right M | P - Pachynolophidae | MP17a | 37.5 | 19.6 | 0.2 | | | | | | | | | | | | | | | |
| 4 | Euzet les Bains | FR | MTM-V.60.1335 | <i>Palaeotherium crassum</i> | right M2(M3?) | P - Palaeotheriidae | MP17a | 37.5 | 20.0 | 0.0 | 20.6 | 1.1 | 10 | 0.7 | | | -8.0 | 0.8 | 6 | 0.7 | -21.7 | | | | |
| 5 | La Débruge | FR | NMB-DB.10 | <i>Palaeotherium magnum</i> | M | P - Palaeotheriidae | MP18 | 36 | 22.5 | 0.1 | | | | | -8.9 | 0.1 | | | | | | -2.8 | 28.0 | 0.1 | 4.0 |
| 5 | La Débruge | FR | NMB-DB.29 | <i>Palaeotherium magnum</i> | l/i | P - Palaeotheriidae | MP18 | 36 | 22.0 | 0.2 | | | | | | | | | | | | | | | |
| 5 | La Débruge | FR | NMB-DB.156 | <i>Anoplotherium sp.</i> | M | A - Anoplotheriidae | MP18 | 36 | 21.2 | 0.2 | 21.9 | 0.7 | 3 | 0.7 | -10.9 | 0.1 | -9.9 | 1.4 | 2 | 2.0 | -23.6 | -2.9 | 27.9 | 0.1 | 4.1 |
| 6 | Gösgen-Canal | CH | NMB-G.C.6 | <i>Palaeotherium curtum</i> | M1 | P - Palaeotheriidae | MP18 | 36 | 19.6 | 0.2 | | | | | | | | | | | | | | | |
| 6 | Gösgen-Canal | CH | NMB-G.C.637 | <i>Palaeotherium curtum</i> | l/i | P - Palaeotheriidae | MP18 | 36 | 19.0 | 0.1 | 19.3 | 0.5 | 2 | 0.7 | | | | | | | | | | | |
| 7 | Radaia (Andrásháza) | RO | MAFI-Ob58- Vt71 | <i>Brachydiastematherium transsylvanicum</i> | left m4 (m3?) | P - Brontotheriidae | MP18 | 36 | 19.2 | 0.0 | 19.2 | 0.3 | 1 | 0.6 | -9.8 | 0.1 | -9.8 | 0.1 | 1 | 0.2 | -23.6 | -5.3 | 25.4 | 0.1 | 5.6 |
| 8 | "Budapest" | HU | MTM-V.60.149 | <i>Amynodon hungaricus</i> | right m2/3 | P - Amynodontidae | ?MP18-19 | 35 | 18.4 | 0.2 | 18.4 | 0.3 | 1 | 0.6 | -8.7 | 0.1 | -8.7 | 0.1 | 1 | 0.2 | -22.5 | -5.4 | 25.3 | 0.1 | 5.8 |
| 2 | Mormont - Entreroches | CH | NMB - no number | Palaeotheriidae indet. | tooth frg. | P - Palaeotheriidae | MP19 | 34.5 | 19.7 | 0.1 | | | | | -9.5 | 0.1 | | | | | | -6.0 | 24.7 | 0.2 | 3.2 |
| 2 | Mormont - Entreroches | CH | NMB - no number | Palaeotheriidae indet. | tooth frg. | P - Palaeotheriidae | MP19 | 34.5 | 20.5 | 0.2 | | | | | -9.5 | 0.1 | | | | | | -4.1 | 26.6 | 0.1 | 4.5 |
| 2 | Mormont - Entreroches | CH | MGL-46784 | <i>Anoplotherium sp.</i> | right P4 | A - Anoplotheriidae | MP19 | 34.5 | 20.6 | 0.1 | | | | | -9.1 | 0.0 | | | | | | -5.0 | 25.8 | 0.1 | 4.5 |
| 2 | Mormont - Éclépens C | CH | MGL-47275 | <i>Palaeotherium crassum renevieri</i> | left M1/2 | P - Palaeotheriidae | MP19 | 34.5 | 21.2 | 0.1 | 20.5 | 0.6 | 4 | 0.6 | | | -9.4 | 0.2 | 3 | 0.2 | -23.2 | | | | |
| 9 | Liptingen 5 | GE | SMNS-no number | <i>Anoplotherium pompeckii</i> | right p4 | A - Anoplotheriidae | MP19 | 34.5 | 17.0 | 0.0 | | | | | -9.6 | 0.1 | | | | | | -8.8 | 21.9 | 0.1 | 3.3 |
| 9 | Liptingen 5 | GE | SMNS-no number | <i>Palaeotherium curtum</i> | left P1 | P - Palaeotheriidae | MP19 | 34.5 | 18.1 | 0.1 | | | | | | | | | | | | | | | |
| 9 | Liptingen 5 | GE | SMNS-no number | ? <i>Anoplotherium sp.</i> | ?l/i3 | A - Anoplotheriidae | MP19 | 34.5 | 20.0 | 0.1 | | | | | | | | | | | | | | | |
| 9 | Liptingen 5 | GE | SMNS-no number | ? <i>Palaeotherium sp.</i> | M3/2, frg. | P - Palaeotheriidae | MP19 | 34.5 | 17.4 | 0.1 | 18.1 | 1.3 | 4 | 1.3 | | | | | | | | | | | |
| 9 | Liptingen 13A | GE | SMNS-no number | <i>Diplobune secundaria</i> | M3 | A - Anoplotheriidae | MP19 | 34.5 | 19.8 | 0.1 | | | | | | | | | | | | | | | |
| 9 | Liptingen 13A | GE | SMNS-no number | <i>Anoplotherium sp.</i> | M?, frg. | A - Anoplotheriidae | MP19 | 34.5 | 18.8 | 0.3 | 19.3 | 0.7 | 2 | 1.0 | | | | | | | | | | | |
| 10 | Neuhausen, Württemberg | GE | MTM-V.60.299 | <i>Palaeotherium minus</i> | M1 | P - Palaeotheriidae | MP19 | 34.5 | 18.3 | 0.1 | | | | | -10.5 | 0.0 | | | | | | -5.5 | 25.2 | 0.1 | 6.6 |
| 10 | Neuhausen, Württemberg | GE | SMNS-4028 | <i>Palaeotherium magnum</i> | right M3 | P - Palaeotheriidae | MP19 | 34.5 | 19.5 | 0.1 | | | | | -10.9 | 0.1 | | | | | | -6.5 | 24.2 | 0.1 | 3.7 |
| 10 | Neuhausen, Württemberg | GE | SMNS-1586 | <i>Palaeotherium muehbergi</i> | left p4 | P - Palaeotheriidae | MP19 | 34.5 | 18.4 | 0.3 | 18.7 | 0.7 | 3 | 0.7 | -9.9 | 0.1 | | | | | | -6.5 | 24.2 | 0.1 | 4.3 |
| 11 | Mähringen | GE | SMNS-no number | <i>Anoplotherium pompeckii</i> | M3 | A - Anoplotheriidae | MP19 | 34.5 | 18.7 | 0.2 | | | | | -9.9 | 0.1 | | | | | | -7.5 | 23.2 | 0.1 | 3.3 |
| 11 | Mähringen | GE | SMNS-1448 | <i>Palaeotherium duvali duvali</i> | left M1 | P - Palaeotheriidae | MP19 | 34.5 | 18.5 | 0.2 | | | | | -10.9 | 0.1 | | | | | | -6.9 | 23.8 | 0.1 | 4.0 |
| 11 | Mähringen | GE | SMNS-no number | <i>Diplobune secundaria</i> | left M3 | A - Anoplotheriidae | MP19 | 34.5 | 19.1 | 0.3 | | | | | -8.4 | 0.0 | | | | | | -5.8 | 24.9 | 0.0 | 6.4 |
| 11 | Mähringen | GE | SMNS-no number | <i>Diplobune quercyi</i> | right M2 | A - Anoplotheriidae | MP19 | 34.5 | 18.4 | 0.1 | 18.7 | 0.3 | 4 | 0.3 | | | | | | | | | | | |
| | | | | | | | | 34.5 | | | 18.6 | 0.9 | 13 | 0.5 | | | -10.0 | 0.9 | 7 | 0.7 | -23.8 | | | | |
| 12 | Frohnstetten | GE | SMNS-1908 | <i>Palaeotherium curtum frohnstettense</i> | left M3 | P - Palaeotheriidae | MP20 | 33.8 | 17.5 | 0.2 | | | | | -12.1 | 0.1 | | | | | | -6.0 | 24.7 | 0.1 | 5.1 |
| 12 | Frohnstetten | GE | SMNS-2534 | <i>Palaeotherium medium suenicum</i> | right P4 | P - Palaeotheriidae | MP20 | 33.8 | 17.1 | 0.2 | | | | | -11.7 | 0.1 | | | | | | -7.2 | 23.5 | 0.1 | 3.3 |
| 12 | Frohnstetten | GE | SMNS-3384 | <i>Plagiolophus fraasi</i> | M3 | P - Pachynolophidae | MP20 | 33.8 | 16.8 | 0.3 | | | | | -11.3 | 0.0 | | | | | | -7.7 | 22.9 | 0.1 | 4.6 |
| 12 | Frohnstetten | GE | SMNS-2852 | <i>Plagiolophus minor</i> | right M3 | P - Pachynolophidae | MP20 | 33.8 | 16.7 | 0.2 | | | | | -11.1 | 0.1 | | | | | | -8.5 | 22.2 | 0.1 | 5.0 |
| 12 | Frohnstetten | GE | SMNS-no number | <i>Anoplotherium pompeckii</i> | M3 | A - Anoplotheriidae | MP20 | 33.8 | 17.2 | 0.1 | 17.0 | 0.3 | 5 | 0.3 | | | -11.5 | 0.4 | 4 | 0.4 | -25.3 | | | | |

ABLE DR2

| Map-Nr. | Locality | Collection & Nr. | Taxon | Sampled material | Family | MAMMAL LEVEL | Age (Ma) | PHOSPHATE | | | | | | CARBONATE IN PHOSPHATE | | | | | | | | | | |
|---------|--------------------------|--------------------|--------------------------------------|------------------|-----------------------|--------------|----------|---------------------------------|-------|------|--------|---|---------|------------------------|------|-------|------|---|---------|-------------------------------------|--------------------------------|---------------------------------|------|----------|
| | | | | | | | | $\delta^{18}\text{O-PO4}$ SMOUW | std.* | AV. | STD.** | N | 1.96xSE | $\delta^{13}\text{C}$ | std. | AV. | STD. | N | 1.96xSE | $\delta^{13}\text{C}_{\text{diet}}$ | $\delta^{18}\text{O-CO3}$ VPDB | $\delta^{18}\text{O-CO3}$ VSMOW | std. | CO3 wt.% |
| 19 | Budapest - Bohn clay pit | HU MTM-V.83.40 | <i>Ranzotherium velaunum</i> | m1/2 | P - Rhinocerotidae | MP23+ | 30.2 | 17.1 | 0.1 | 17.1 | 0.3 | 1 | 0.6 | -11.1 | 0.1 | -11.1 | 0.1 | 1 | 0.2 | -24.8 | -7.0 | 23.6 | 0.1 | 5.5 |
| 19 | Budapest - Ujlak | HU MTM-V.60.308 | ? <i>Eggysodon</i> sp. | right p3? | P - Hyracodontidae | MP23-24 | 30 | 16.2 | 0.2 | 16.2 | 0.3 | 1 | 0.6 | -13.6 | 0.1 | -13.6 | 0.1 | 1 | 0.2 | -27.3 | -8.3 | 22.4 | 0.0 | 3.3 |
| 16 | Cluj | RO UBB-no number | <i>Elomeryx borbonicus</i> | P? | A - Anthracotheriidae | MP23-24 | 30 | 16.9 | 0.1 | | | | | | | | | | | | | | | |
| 16 | Cluj | RO UBB-no number | <i>Elomeryx borbonicus</i> | I | A - Anthracotheriidae | MP23-24 | 30 | 16.8 | 0.1 | 16.9 | 0.1 | 2 | 0.1 | | | | | | | | | | | |
| 20 | Aghires | RO UBB-no number | ? <i>Anthracotherium monsiense</i> | | A - Anthracotheriidae | MP23-24 | 30 | 15.2 | 0.1 | 15.2 | 0.3 | 1 | 0.6 | | | | | | | | | | | |
| | | | | | | | 30 | | | 16.3 | 1.0 | 3 | 1.1 | | | | | | | | | | | |
| 21 | Csákberény | HU MTM-V.60.284 | <i>Protacatherium albigenae</i> | right M2 frg. | P - Rhinocerotidae | MP23-25 | 29.4 | 16.0 | 0.0 | | | | | -13.3 | 0.1 | | | | | | -6.9 | 23.8 | 0.1 | 5.9 |
| 21 | Csákberény | HU MTM-V.60.289 | <i>Protacatherium albigenae</i> | right M2 frg. | P - Rhinocerotidae | MP23-25 | 29.4 | 16.6 | 0.2 | 16.3 | 0.4 | 2 | 0.6 | -12.8 | 0.1 | -13.1 | 0.3 | 2 | 0.5 | -26.8 | -7.0 | 23.7 | 0.1 | 3.2 |
| 22 | Poillat | CH MJSN-POI007-346 | <i>Ranzotherium</i> | tooth frg. | P - Rhinocerotidae | MP24- | 29.6 | 17.6 | 0.3 | | | | | -9.2 | 0.1 | | | | | | -4.8 | 26.0 | 0.1 | |
| 22 | Poillat | CH MJSN-POI007-236 | <i>Ranzotherium</i> | tooth frg. | P - Rhinocerotidae | MP24- | 29.6 | 19.9 | 0.3 | | | | | -10.0 | 0.1 | | | | | | -4.2 | 26.5 | 0.1 | |
| 22 | Poillat | CH MJSN-POI007-188 | <i>Ranzotherium</i> | tooth frg. | P - Rhinocerotidae | MP24- | 29.6 | 18.7 | 0.3 | | | | | -10.3 | 0.1 | | | | | | -3.1 | 27.7 | 0.1 | |
| 22 | Poillat | CH MJSN-POI007-211 | <i>Ranzotherium</i> | tooth frg. | P - Rhinocerotidae | MP24- | 29.6 | 16.8 | 0.3 | 18.3 | 1.3 | 4 | 1.3 | -10.0 | 0.1 | -9.9 | 0.5 | 4 | 0.5 | -23.6 | -6.6 | 24.1 | 0.1 | |
| 23 | Tatabánya - Felsőgalla | HU MTM-V.60.154 | ? <i>Ranzotherium cf. romani</i> * | left P2 | P - Rhinocerotidae | ?MP24-25 | 28.6 | 14.6 | 0.2 | | | | | -12.0 | 0.1 | | | | | | -8.2 | 22.4 | 0.2 | 3.9 |
| 23 | Tatabánya - Felsőgalla | HU MTM-V.60.192 | <i>Ranzotherium cf. romani</i> * | left m2/3 | P - Rhinocerotidae | MP24-25 | 28.6 | 14.9 | 0.1 | | | | | -12.3 | 0.1 | | | | | | -8.1 | 22.5 | 0.1 | 4.7 |
| 23 | Tatabánya - Felsőgalla | HU MTM-V.60.590 | ? <i>Anthracotherium monsiense</i> * | M1/2 frg. | A - Anthracotheriidae | MP24-25 | 28.6 | 16.0 | 0.0 | | | | | -11.3 | 0.1 | | | | | | -7.6 | 23.0 | 0.2 | 4.6 |
| 23 | Tatabánya - Felsőgalla | HU MTM-V.60.1569 | ? <i>Anthracotherium monsiense</i> * | C/c frg. | A - Anthracotheriidae | MP24-25 | 28.6 | 15.1 | 0.1 | 15.2 | 0.6 | 4 | 0.6 | -11.2 | 0.0 | -11.7 | 0.5 | 4 | 0.5 | -25.5 | -8.3 | 22.4 | 0.1 | 9.3 |
| 24 | Bumbach | CH NMB-UM7155 | <i>Anthracotherium bumbachense</i> | i | A - Anthracotheriidae | MP25- | 28 | 16.1 | 0.2 | | | | | -10.0 | 0.0 | | | | | | -8.4 | 22.2 | 0.1 | 3.2 |
| 24 | Bumbach | CH NMB-UM13 | <i>Ranzotherium</i> sp. | p | P - Rhinocerotidae | MP25- | 28 | 15.6 | 0.4 | | | | | -11.3 | 0.1 | | | | | | -8.2 | 22.5 | 0.1 | 4.2 |
| 24 | Bumbach | CH NMB-UM6279 | <i>Ranzotherium</i> sp. | p | P - Rhinocerotidae | MP25- | 28 | 16.7 | 0.3 | 16.1 | 0.6 | 3 | 0.6 | -9.9 | 0.0 | -10.4 | 0.8 | 3 | 0.8 | -24.2 | -7.8 | 22.8 | 0.1 | 3.3 |

CH - Switzerland
 GE - Germany
 FR - France
 HU - Hungary
 IT - Italy
 RO - Romania

* DB and LS new classification here (MP levels too)

P - Perissodactyla, A - Artiodactyla

* standard deviation of triplicate analyses
 **for single sample analyses 0.3 std is used
 derived from the max. error on standard calibration

Preparation of the samples and their stable isotope analyses

The surface of the teeth was always cleaned before sampling, then tooth enamel was carefully shaven off by a micro-drill, while bigger enamel pieces were powdered and homogenized in an agate mortar. The sample powders were pre-treated in two steps (Koch et al., 1997): 1) leached in 2.5% NaOCl for 24 hours to remove any possible residual organic matter; 2) a wash in 1N acetic acid-Ca-acetate (pH=4.5, 6 hours) to eliminate any exogenous carbonates. Between and after these steps the samples were rinsed several times in distilled water. NBS-120c phosphorite reference material was always prepared parallel with each batch of sample set.

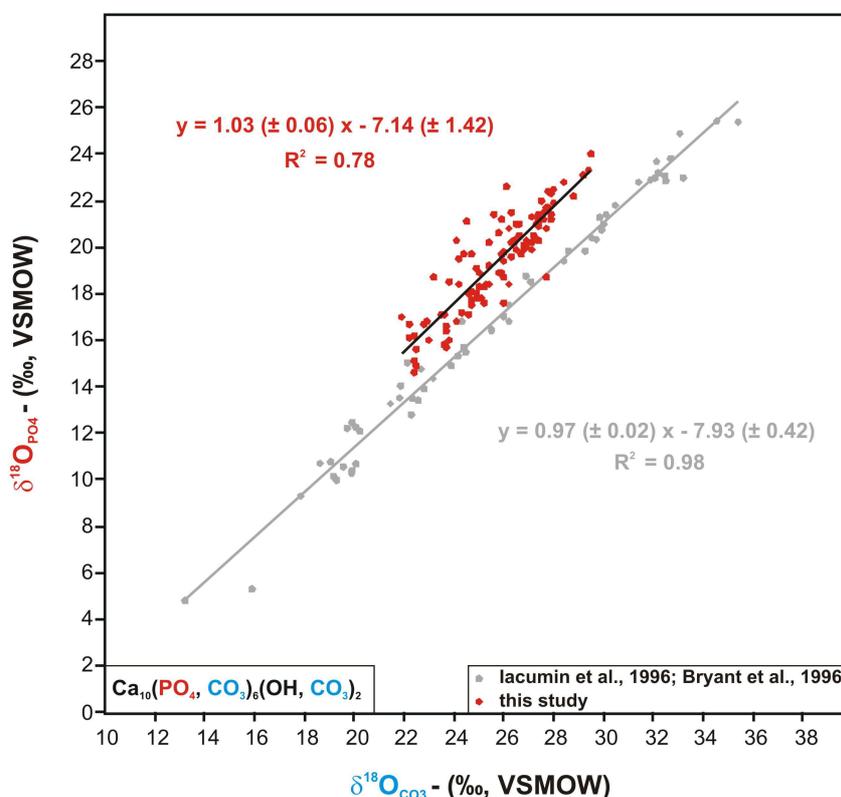
The oxygen and carbon isotopic compositions of structural carbonate in phosphate [$\delta^{18}\text{O}_{\text{CO}_3}$, $\delta^{13}\text{C}$] were directly analyzed on the pre-cleaned sample powders using a Gasbench II coupled to a Finnigan MAT Delta Plus XL mass spectrometer. The measured isotopic ratios were normalized to an in-house Carrara marble calcite standard that is calibrated against NBS-19. The analytical precision for this method is better than $\pm 0.1\text{‰}$ for O and C isotopes (e.g., Spötl and Vennemann, 2003). The $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ values are expressed in δ -notation relative to VPDB (Vienna Pee Dee Belemnite).

The phosphate oxygen isotopic composition [$\delta^{18}\text{O}_{\text{PO}_4}$] The PO_4^{3-} group was further separated from other oxygen bearing ions in the apatite structure (CO_3^{2-} , OH), applying a silver phosphate precipitation method (O'Neil et al., 1994; Dettman et al., 2001; Kocsis, 2011). The $\delta^{18}\text{O}_{\text{PO}_4}$ was analyzed on a TC/EA (high-temperature conversion elemental analyzer) (Vennemann et al., 2002) coupled to a Finnigan MAT Delta Plus XL mass spectrometer, where the silver-phosphate is converted to CO at 1450°C via reduction with graphite. The results were corrected to in-house Ag_3PO_4 phosphate standards (LK-2L: 12.1‰ and LK-3L: 17.9‰) that had better than $\pm 0.3\text{‰}$ (1σ) standards deviations during measurements. These Ag_3PO_4 phosphate standards were calibrated by TC/EA to TU-1 and TU-2 standards using values defined by conventional fluorination method (CF)

(cf. [Vennemann et al., 2002](#)) and were also calibrated with laser-fluorination method yielding identical values.

For the NBS-120c phosphorite reference material an average values of $21.6 \pm 0.3\text{‰}$ ($n=21$) was obtained. Oxygen isotope compositions are expressed in the δ -notation relative to Vienna Standard Mean Ocean Water (VSMOW). All the stable isotope analyses were carried out in the stable isotope laboratory of the Institute of Earth Sciences at University of Lausanne, Switzerland.

Figure DR1. Phosphate and structural carbonate oxygen isotopic compositions in bioapatite



The grey dots and grey regression line are derived from datasets of modern mammals ([Iacumin et al., 1996](#); [Bryant et al., 1996](#)). Note that the late Eocene-early Oligocene samples show parallel regression line with significant correlation and with similar slope (a) and slightly higher intercept (b) to the modern mammals. Considering the standard errors (SE) on both (a) and (b), the 95% confidential intervals cover the parameters of the modern dataset ($a \pm 0.11$; $b \pm 2.79$). Moreover, assuming that the modern data represent the real correlation between $\delta^{18}\text{O}_{\text{CO}_3}$ and $\delta^{18}\text{O}_{\text{PO}_4}$ of the whole population (statistically speaking from which the fossil sub-samples may have been taken), then a simply Z-test on the SE of the slope and the intercept in the fossil dataset yield no significant

differences from the mean of these parameters of the modern population (95% conf. limit, i.e. $1.96 \times \text{SE}$). All these suggest that the fossil teeth are well-preserved and no significant diagenetic isotopic alteration occurred in tooth enamel.

Further notes: The $\delta^{18}\text{O}_{\text{CO}_3}$ is chosen as the “independent” variable (x) because these values have much smaller error associated with them concerning their measurements when compared to $\delta^{18}\text{O}_{\text{PO}_4}$ analyses. The apparently more disperse fossil dataset may perhaps link to minor alteration in $\delta^{18}\text{O}_{\text{CO}_3}$, hence the reason for using $\delta^{18}\text{O}_{\text{PO}_4}$ in this work to investigate *in-vivo* environmental factors (see also Kohn and Cerling, 2002; Zazzo et al., 2004 cited in the paper).

Table DR3. t-tests on the major taxa at four localities

| Map-Nr. | Locality | Taxon | Family | MAMMAL LEVEL | Age (Ma) | $\delta^{18}\text{O-PO4}$ SMOW | std. | $\delta^{13}\text{C}$ | std. | | |
|-----------------------|-------------|------------------------------------------------------------------|-----------------------|--------------|----------|--------------------------------|------|-----------------------|------|---------------------|-----------------------|
| Artiodactyla | | | | | | | | | | | |
| 15 | Möhren 19 | GE <i>Diplobune</i> sp. | A - Anoplotheriidae | MP21 | 33.25 | 21.4 | 0.1 | -11.1 | 0.0 | t(8)= 0.42, p=0.69 | $\delta^{18}\text{O}$ |
| 15 | Möhren 19 | GE <i>Diplobune</i> cf. <i>quercyi</i> | A - Anoplotheriidae | MP21 | 33.25 | 18.9 | 0.1 | | | t(7)= 1.02, p=0.34 | $\delta^{13}\text{C}$ |
| 15 | Möhren 19 | GE <i>Diplobune</i> cf. <i>quercyi</i> | A - Anoplotheriidae | MP21 | 33.25 | 18.4 | 0.1 | -12.2 | 0.1 | | |
| 15 | Möhren 19 | GE <i>Diplobune</i> sp. | A - Anoplotheriidae | MP21 | 33.25 | 20.2 | 0.3 | -10.7 | 0.0 | | |
| 15 | Möhren 19 | GE <i>Anoplotherium commune</i> | A - Anoplotheriidae | MP21 | 33.25 | 18.1 | 0.1 | -14.1 | 0.1 | | |
| 15 | Möhren 19 | GE <i>Anoplotherium commune</i> | A - Anoplotheriidae | MP21 | 33.25 | 17.1 | 0.1 | -13.2 | 0.1 | | |
| Perissodactyla | | | | | | | | | | | |
| 15 | Möhren 19 | GE <i>Palaeotherium medium</i> | P - Palaeotheriidae | MP21 | 33.25 | 17.8 | 0.1 | -12.1 | 0.1 | | |
| 15 | Möhren 19 | GE <i>Palaeotherium medium</i> | P - Palaeotheriidae | MP21 | 33.25 | 17.7 | 0.0 | -11.4 | 0.1 | | |
| 15 | Möhren 19 | GE <i>Palaeotherium medium</i> | P - Palaeotheriidae | MP21 | 33.25 | 18.0 | 0.1 | -11.9 | 0.1 | | |
| 15 | Möhren 19 | GE <i>Palaeotherium medium</i> | P - Palaeotheriidae | MP21 | 33.25 | 20.9 | 0.0 | -10.4 | 0.1 | | |
| Artiodactyla | | | | | | | | | | | |
| 15 | Möhren13 | GE <i>Elomeryx crispus crispus</i> | A - Anthracotheriidae | MP22 | 31.7 | 17.5 | 0.3 | | | t(13)= 1.78, p=0.10 | $\delta^{18}\text{O}$ |
| 15 | Möhren13 | GE <i>Elomeryx crispus crispus</i> | A - Anthracotheriidae | MP22 | 31.7 | 16.4 | 0.3 | | | t(10)= 1.54, p=0.16 | $\delta^{13}\text{C}$ |
| 15 | Möhren13 | GE <i>Elomeryx crispus crispus</i> | A - Anthracotheriidae | MP22 | 31.7 | 15.7 | 0.1 | -11.1 | 0.1 | | |
| 15 | Möhren13 | GE <i>Elomeryx crispus crispus</i> | A - Anthracotheriidae | MP22 | 31.7 | 16.4 | 0.1 | -11.7 | 0.1 | | |
| 15 | Möhren13 | GE <i>Elomeryx crispus crispus</i> | A - Anthracotheriidae | MP22 | 31.7 | 17.2 | 0.0 | -11.0 | 0.0 | | |
| 15 | Möhren13 | GE <i>Elomeryx crispus crispus</i> | A - Anthracotheriidae | MP22 | 31.7 | 15.8 | 0.1 | -10.7 | 0.0 | | |
| 15 | Möhren13 | GE <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP22 | 31.7 | 18.3 | 0.2 | -11.3 | 0.1 | | |
| 15 | Möhren13 | GE <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP22 | 31.7 | 18.9 | 0.0 | -11.4 | 0.1 | | |
| 15 | Möhren13 | GE <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP22 | 31.7 | 19.7 | 0.2 | -12.5 | 0.1 | | |
| Perissodactyla | | | | | | | | | | | |
| 15 | Möhren13 | GE <i>Epiacceratherium magnum</i> | P - Rhinocerotidae | MP22 | 31.7 | 18.1 | 0.1 | | | | |
| 15 | Möhren13 | GE <i>Epiacceratherium magnum</i> | P - Rhinocerotidae | MP22 | 31.7 | 19.7 | 0.1 | -11.7 | 0.0 | | |
| 15 | Möhren13 | GE <i>Epiacceratherium sp.</i> | P - Rhinocerotidae | MP22 | 31.7 | 17.8 | 0.2 | -12.3 | 0.1 | | |
| 15 | Möhren13 | GE <i>Epiacceratherium magnum</i> | P - Rhinocerotidae | MP22 | 31.7 | 18.0 | 0.2 | -12.0 | 0.1 | | |
| 15 | Möhren13 | GE <i>Epiacceratherium magnum</i> | P - Rhinocerotidae | MP22 | 31.7 | 18.7 | 0.2 | -12.1 | 0.1 | | |
| 15 | Möhren13 | GE <i>Epiacceratherium magnum</i> | P - Rhinocerotidae | MP22 | 31.7 | 18.3 | 0.2 | -11.2 | 0.0 | | |
| Artiodactyla | | | | | | | | | | | |
| 17 | Villebramar | FR <i>Entelodon</i> sp. | A - Entelodontidae | MP22 | 31.7 | 19.4 | 0.2 | -11.2 | 0.0 | t(12)= 0.04, p=0.97 | $\delta^{18}\text{O}$ |
| 17 | Villebramar | FR <i>Entelodon</i> sp. | A - Entelodontidae | MP22 | 31.7 | 18.9 | 0.3 | -10.5 | 0.1 | t(12)= 0.88, p=0.23 | $\delta^{13}\text{C}$ |
| 17 | Villebramar | FR <i>Entelodon</i> sp. | A - Entelodontidae | MP22 | 31.7 | 23.1 | 0.2 | -10.7 | 0.1 | | |
| Perissodactyla | | | | | | | | | | | |
| 17 | Villebramar | FR <i>Plagiolophus</i> sp. | P - Pachynolophidae | MP22 | 31.7 | 21.9 | 0.2 | -10.6 | 0.1 | | |
| 17 | Villebramar | FR <i>Plagiolophus</i> sp. | P - Pachynolophidae | MP22 | 31.7 | 21.5 | 0.2 | -10.0 | 0.1 | | |
| 17 | Villebramar | FR <i>Plagiolophus</i> sp. | P - Pachynolophidae | MP22 | 31.7 | 24.0 | 0.2 | -9.9 | 0.0 | | |
| 17 | Villebramar | FR <i>Plagiolophus</i> sp. | P - Pachynolophidae | MP22 | 31.7 | 23.3 | 0.2 | -9.8 | 0.1 | | |
| 17 | Villebramar | FR <i>Plagiolophus</i> sp. | P - Pachynolophidae | MP22 | 31.7 | 21.4 | 0.2 | -11.0 | 0.0 | | |
| 17 | Villebramar | FR <i>Ronzotherium</i> sp. | P - Rhinocerotidae | MP22 | 31.7 | 18.4 | 0.3 | -10.8 | 0.1 | | |
| 17 | Villebramar | FR <i>Ronzotherium</i> sp. | P - Rhinocerotidae | MP22 | 31.7 | 18.9 | 0.0 | -10.5 | 0.1 | | |
| 17 | Villebramar | FR <i>Ronzotherium</i> sp. | P - Rhinocerotidae | MP22 | 31.7 | 20.1 | 0.2 | -11.4 | 0.1 | | |
| 17 | Villebramar | FR <i>Ronzotherium</i> sp. | P - Rhinocerotidae | MP22 | 31.7 | 18.4 | 0.2 | -8.8 | 0.1 | | |
| 17 | Villebramar | FR <i>Ronzotherium</i> sp. | P - Rhinocerotidae | MP22 | 31.7 | 20.2 | 0.2 | -10.4 | 0.1 | | |
| 17 | Villebramar | FR <i>Ronzotherium</i> sp. | P - Rhinocerotidae | MP22 | 31.7 | 17.6 | 0.1 | -11.2 | 0.1 | | |
| Artiodactyla | | | | | | | | | | | |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.0 | 0.1 | -9.8 | 0.1 | t(14)= 1.45, p=0.17 | $\delta^{18}\text{O}$ |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.8 | 0.2 | -8.5 | 0.1 | t(10)= 2.01, p=0.07 | $\delta^{13}\text{C}$ |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 21.7 | 0.1 | -8.3 | 0.1 | | |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 21.2 | 0.1 | | | | |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 22.2 | 0.1 | -7.7 | 0.0 | | |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.2 | 0.1 | -9.3 | 0.1 | | |
| 13 | Monteviale | IT <i>Anthracotherium monsvialense</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.3 | 0.3 | -8.2 | 0.0 | | |
| 13 | Monteviale | IT <i>Anthracochoeerus stehlini</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.0 | 0.2 | | | | |
| 13 | Monteviale | IT <i>Anthracochoeerus stehlini</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.1 | 0.1 | | | | |
| 13 | Monteviale | IT <i>Anthracochoeerus stehlini</i> | A - Anthracotheriidae | MP21- | 33.4 | 20.7 | 0.3 | | | | |
| Perissodactyla | | | | | | | | | | | |
| 13 | Monteviale | IT <i>Epiacceratherium bolcense</i> (<i>Trigonias ombonii</i>) | P - Rhinocerotidae | MP21- | 33.4 | 21.0 | 0.2 | -9.0 | 0.0 | | |
| 13 | Monteviale | IT <i>Epiacceratherium bolcense</i> (<i>Trigonias ombonii</i>) | P - Rhinocerotidae | MP21- | 33.4 | 20.5 | 0.0 | -9.1 | 0.1 | | |
| 13 | Monteviale | IT <i>Epiacceratherium bolcense</i> (<i>Trigonias ombonii</i>) | P - Rhinocerotidae | MP21- | 33.4 | 21.2 | 0.2 | -8.9 | 0.1 | | |
| 13 | Monteviale | IT <i>Epiacceratherium bolcense</i> (<i>Trigonias ombonii</i>) | P - Rhinocerotidae | MP21- | 33.4 | 20.3 | 0.2 | -9.8 | 0.1 | | |
| 13 | Monteviale | IT <i>Epiacceratherium bolcense</i> (<i>Trigonias ombonii</i>) | P - Rhinocerotidae | MP21- | 33.4 | 22.4 | 0.2 | -9.5 | 0.1 | | |
| 13 | Monteviale | IT <i>Epiacceratherium bolcense</i> (<i>Trigonias ombonii</i>) | P - Rhinocerotidae | MP21- | 33.4 | 22.8 | 0.3 | -9.8 | 0.1 | | |

Figure DR2. Detailed paleogeographic maps (from Figure 3)

The paleomaps are based on compilation of data from many different works. See reference list for details.

Abbreviations on the maps:

AMB – Alpine Molasse Basin, **Ap** – Apulia, **Aq** – Aquitanian basin, **BM** – Bohemian Massif, **Ca** – Calabria, **CF** – Carpathian Foreland, **Da** – Dacides, **EA** – Estern Alps, **EB** – Ebro Basin, **ED** – External Dinarides, **IM** – Iberian Massif, **MC** – Massif Central, **Mo** – Moesian, **NSB** – North Sea Basin, **PB** – Paris Basin, **PL** – Polish Lowlands, **Rh** – Rhodopes, **RhG** – Rhine Graben, **SA** – Southern Alps, **SG** – Supra Getic, **SM** – Serbo-Macedonian Massif, **SZ** – Sava Zone, **Td** – Transdanubian, **ThB** – Thrace Basin, **Ti** – Tisza unit, **Ts** – Transylvanian Paleogene Basin, **VH** – Volhynian High, **WA** – Western Alps, **WCp** – Western Carpathians.

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