

Methods

REE contents in biogenic apatite were obtained by instrumental neutron activation analysis (INAA) in the Institute of Precambrian Geology and Geochronology, St. Petersburg. Handpicked and carefully purified air-dried samples of 0.3–15 mg were placed in pure quartz vials and irradiated for 100 hr with epithermal neutrons at the Institute of Nuclear Physics, St. Petersburg (Reactor VVR-1). The density of neutron flux is 5×10^{18} neutrons/cm²s⁻¹. The measurement procedure was described by Felitsyn et al. (1990), mean relative errors are less than 3% for Sm, Nd, and Tb, less than 5% for La, Ce, Ho, and less than 10% for Nd and Tb.

For Sm-Nd determination crushed samples were spiked by ¹⁴⁶Nd – ¹⁴⁹Sm and decomposed during 2-5 days in an oven at 120 °C. Sm and Nd separation was carried out according to standard methods of two-stage ion-exchange and extraction chromatography. The measurements were performed on a Finnegan MAT-261 mass spectrometer equipped with eight collectors under static mode at the Institute of Precambrian Geology and Geochronology, St. Petersburg.

The ¹⁴³Nd/¹⁴⁴Nd ratio was normalized within run to ¹⁴⁸Nd/¹⁴⁴Nd = 0.241570 and then adjusted to a ¹⁴³Nd/¹⁴⁴Nd value of 0.511860 (La Jolla standard). Assigned errors (2σ) for ¹⁴⁷Sm/¹⁴⁴Nd and ¹⁴³Nd/¹⁴⁴Nd were $\pm 0.3\%$ and ± 0.000015 of standard (external reproducibility). The 2σ errors cited in table for ¹⁴³Nd/¹⁴⁴Nd reflect in-run precision and demonstrate only the quality of these analyses. The blank level for Sm was 0.01 ng and was for Nd 0.05 ng. The data obtained for BCR-1 during the course of this analytical work are: Sm = 6.487 ppm, Nd = 28.45 ppm, ¹⁴³Nd/¹⁴⁴Nd = 0.512663 ± 0.000009 , ¹⁴⁷Sm/¹⁴⁴Nd = 0.13829 (for details, see Ovchinnikova et al. 1995).

Nd isotopic analyses are reported in the standard epsilon notation where $\epsilon_{Nd}(T) = [(^{143}Nd/^{144}Nd)_{sample}(t)/(^{143}Nd/^{144}Nd)_{CHUR}(t)-1]10^4$, where CHUR is

the chondritic uniform reservoir with a present-day $^{143}\text{Nd}/^{144}\text{Nd} = 0.512638$ and $^{147}\text{Sm}/^{144}\text{Nd} = 0.1967$.

REFERENCES CITED

Felitsyn, S. B., Vaganov, P. A., and Kirianov, V. Y., 1990, Distribution of rare and trace elements in ashes of Kamchatka volcanoes based on instrumental neutron activation analysis: Volcanology and Seismology, v. 2, p. 23-35.

Ovchinnikova, G. V., Belyatsky, B. V., Vasilieva, I. M., and Levsky, L. K., 1995, Sr-Nd Pb isotope characteristics of the mantle sources of basalts from the Canary Islands: Petrology, v. 3(2), p. 172-182.

TABLE A

Sample	Material	Age Biozones	Sm ppm	Nd ppm	147Sm/144Nd	143Nd/144Nd +/-2s measured. ¹	Host rock type
Ma-U1	brachiopod	A. <i>Pisiformis/Homognostus</i>	27.39	126.4	0.13149	0.521116+/-13	sandstone
M-77/9	brachiopod	<i>Peltura</i>	71.84	290.2	0.15013	0.512062+/-9	sandstone
T-3/28	brachiopod	<i>Cordyliodus proavus</i> (redepos.)	59.88	275.6	0.13178	0.512051+/-7	sandstone
Syas-1	brachiopod	<i>Cordyliodus proavus</i>	68.51	297.9	0.13947	0.512124+/-9	sandstone
Lava 1	brachiopod	<i>Cordyliodus proavus</i>	54.96	239.1	0.13937	0.512112+/-27	sandstone
U83-8/9	brachiopod	<i>Cordyliodus lindstromi</i>	48.92	223.96	0.13246	0.512036+/-6	sandstone
U-83-6	brachiopod	<i>Cordyliodus proavus</i>	34.55	155.3	0.13494	0.512033+/-46	sandstone
MB I beta	conodont	<i>Prioniodus elegans</i>	97.78	425.6	0.13933	0.512149+/-13	limestone
MB IIab	conodont	<i>Oepliodus evae</i>	149.9	661.1	0.13668	0.5121224+/-16	limestone
Tonu-5	brachiopod	<i>Baltioniodus navis/B. triangularis</i>	250	1079	0.14045	0.512165+/-8	limestone
Tonu-24	brachiopod	<i>Paroistodus originalis</i>	189.7	957.2	0.1342	0.512129+/-12	limestone
Tonu-30	brachiopod	<i>Microzarkofina flabellum parva</i>	207.9	899.1	0.14024	0.512219+/-6	limestone
KJ I beta	brachiopod	<i>Prioniodus elegans</i>	199.3	881	0.13718	0.512036+/-7	limestone
6817-8C	brachiopod	<i>Oepliodus evae</i>	355.1	1375	0.15665	0.512295+/-5	limestone
EO-2	biogen. apatite unidentified	<i>Eoplacognathus variabilis</i>	107.6	461	0.14155	0.512225+/-5	limestone
E22-2	biogen. apatite unidentified	<i>Eoplacognathus variabilis</i>	218.1	842.1	15703	0.512281+/-11	limestone
E11-5	biogen. apatite unidentified	<i>Eoplacognathus variabilis</i>	394.8	1475	0.1623	0.512106+/-5	limestone
R11-5	biogen. apatite unidentified	<i>Eoplacognathus variabilis</i>	363.9	1415	0.15592	0.512214+/-5	limestone
E21-4	biogen. apatite unidentified	<i>Eoplacognathus variabilis</i>	118.3	470.3	0.15269	0.512109+/-11	limestone
Hallek	biogen. apatite unidentified	<i>Eoplacognathus variabilis</i>	92.05	399	0.13993	0.512199+/-8	limestone
SV-1	volcanic ash, residue	<i>Eoplacognathus suecicus</i>	7.78	39.16	0.12046	0.512244+/-6	limestone
225555	phosphonite, Greenland	Middle Cambrian, Greenland	18.5	90.1	0.1245	0.511990+/-7	
225556	phosphonite, Greenland	Middle Cambrian, Greenland	15.46	76.17	0.12306	0.512006+/-8	
225569	phosphonite, Greenland	Middle Cambrian, Greenland	20.76	100.2	0.1257	0.511995+/-6	
VFS-Ph	phosphonite, Moscow syneclide	Early Cambrian Rovno	95.4	542	0.12308	0.511915+/-9	
VTyR	phosphonite, Moscow syneclide	Early Cambrian Rovno	108.4	606	0.13377	0.511985+/-4	
2 BS	Shale, Moscow syneclide	Vendian Redkino	6.3	36.1	0.10562	0.511105+/-13	
8 BS	Shale, Moscow syneclide	Vendian Kottlin	6.84	39.11	0.10612	0.511836+/-13	
37 BS	Shale, Moscow syneclide	Cambrian Rovno	4.23	21.77	0.11786	0.511890+/-8	

TABLE B

	La	116	172	160	210	241	158	165	367	701	586	519	681	393
	Ce	270	577	414	660	760	397	456	1298	1722	1425	1559	1671	842
	Pr	Nd	140	401	260	351	330	238	260	795	1368	1089	864	1225
Sm	30	73	47	63	76	43	48	148	264	198	195	256	128	
Eu	7.9	14.6	10.3	16.7	19.7	10.8	13.2	30.8	54.2	40.7	44	58	28	
Gd														
Tb	6.5	11.9	9	11	13	9.2	9.9	21	36	27	31	258	126	
Dy														
Ho	8	10.5	10	12	17	11	12.2	18.8	35.1	29.5				
Er														
Tm														
Yb	10.4	14	13.6	17.6	23.6	14.2	15.4	25.4	46.1	43.6	31	34.4	27.1	
Lu														

Sample nr.: **Ma-U1** **T-3/28** **E-10/2** **Syas-1** **Lava-1** **U-83-6** **U-83-8/9** **M/Bibeta** **Tonu-5** **Tonu-24** **EST-1** **EST-2** **EST-3**

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Conodont zones *A. pisiformis* *Rredepo.* in *C. andresi* *C. proavus* *C. proavus* *C. proavus* *C. proavus* *C. proavus* *C. elegans* *Hornagnost. Cordyloodus* *Paroliododus* *Frixiolodus* *linostromi* *elegans* *navigatrix* *originalis*