

Supplemental Material

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1 MATERIALS AND METHODS

2 Field Methods

3 A plastic trowel was used to sample soil where exposed between plants within 0.5 cm of the
4 surface. Voids between marbles in dust trap allows dust particles to fall or to be washed into a 2/
5 polycarbonate bottle attached to the spout of the funnel (Fig. SI-1), thus eliminating the
6 possibility of re-entrainment of particles.

8 Analytical Procedures

9 A mixture of 9.5 ml of 18 MΩ D.I. water and 1.0 ml of 70% distilled HNO₃ was added to 0.3-0.7
10 g of soil. Samples then sat in a 65° C hot bath for 30 minutes, sonicated for 15 minutes, shaken
11 for 12 hours, and then chilled to 4° C for 24 h. Digested solutions were then warmed to room
12 temperature for 1 h before diluting 0.2 ml of particle-free supernatant with 10 ml of diluting
13 solution, which contained internal standards Rh, In, and Bi. Reproducibility of sample
14 homogenization and digestion was verified by analyzing soils digested in triplicate every 10-15
15 samples (n=36), and supernatant from the three digestions were also analyzed in triplicate. Most
16 coefficients of variation (CV) for samples analyzed in triplicate are between 1 and 7% for metals
17 of interest; the average CV for all 36 samples analyzed in triplicate is 9%, which is sufficient for
18 this study (Atteia et al., 1994). However, even with thorough homogenization, 9 samples tested
19 in triplicate yielded larger CVs for Pb, which may be attributable to strong Pb inhomogeneities
20 among soil particles within individual samples. For plant samples, 0.01-0.05 g of root and shoot
21 material was placed in a Teflon digestion vessel with 1 ml of HNO₃ (Jones, 2001). Digestion
22 vessels were transferred to an 80° C hot bath for 1 hr, cooled to room temperature, and then 1 ml
23 of 35% H₂O₂ was added and vessels were transferred back to the hot bath for one hour (Jones,

24 2001). Once digestions cooled to room temperature, 0.2 ml of supernatant was diluted with 10
25 ml of diluting solution for analysis by ICP-MS (Table SI3). Blanks and standards were analyzed
26 at regular intervals throughout sample runs. Instrumentation accuracy for soil and plant analyses
27 was verified with certified reference materials PACS-2 (marine sediment) and ERM-CD281 (rye
28 grass), respectively.



Figure SII. One of four dust traps deployed on the Junin Plain between June 2016 and June 2017. Polycarbonate traps are 43 cm in diameter and filled with 6 cm of glass marbles on a nylon mesh following design described by Reheis and Kihl (1995). Two-liter polycarbonate bottle collects sediment when washed through marbles.

Table SI-2.

Metal Concentrations in Soil Profile (Fig. 2) at Site 2 (Fig. 3)

Sample ID	Depth (cm)	Al (mg/kg)	Zn (mg/kg)	Cu (mg/kg)	As (mg/kg)	Ag (mg/kg)	Pb (mg/kg)
SP1-1	0	8545.087	47.766	72.271	70.798	0.819	181.292
SP1-2	1	12381.838	86.310	33.381	81.417	0.540	181.268
SP1-3	2	12650.609	89.014	30.021	72.549	0.513	181.033
SP1-4	4	11893.875	56.630	22.448	15.722	0.204	104.585
SP1-5	6	13057.251	9.794	20.936	1.611	0.037	29.146
SP1-6	8	13641.646	6.756	20.141	1.146	0.019	23.098
SP1-7	10	14376.657	7.097	21.775	1.531	0.023	25.063
SP1-8	14	22648.602	8.315	32.095	2.093	0.025	38.350
SP1-9	18	14249.304	5.164	19.915	1.761	0.018	23.022
SP1-10	22	13803.333	4.348	19.151	2.006	0.025	22.047
SP1-11	30	14532.698	4.446	19.018	2.957	0.021	23.463
SP1-12	35	12260.225	17.509	3.527	4.665	0.020	23.890

Table SI-3.
Metal Concentrations in Plant Roots and Shoots, and Adjacent Soil

Soil Sample Site	latitude (°)	longitude (°)	distance from mining district (km)	azimuth bearing from mining district (°)	lab code	material	factor	Al (mg/kg)	Cu (mg/kg)	Zn (mg/kg)	As (mg/kg)	Ag (mg/kg)	Pb (mg/kg)
H6	-10.8083	-76.2865	14.7	190.9	H6R	roots		4886.860	87.909	99.548	17.933	0.697	81.235
					H6C	shoots		352.080	21.542	32.332	3.049	0.437	15.390
					H6	soil		11779.283	221.250	338.805	55.233	1.713	633.136
							<i>translocation factor</i>	7%	25%	32%	17%	63%	19%
							<i>accumulation factor</i>	22%	25%	19%	19%	33%	8%
G20	-10.9762	-76.0217	42.2	141.8	G20R	roots		998.745	7.532	53.272	3.272	1.179	12.419
					G20C	shoots		460.466	3.852	26.599	1.198	0.068	4.685
					G20	soil		8119.405	13.192	56.554	38.380	0.083	107.301
							<i>translocation factor</i>	46%	51%	50%	37%	6%	38%
							<i>accumulation factor</i>	9%	43%	71%	6%	750%	8%
H4	-10.8182	-76.2809	15.7	187.8	H4R	roots		1080.491	53.220	666.773	11.443	1.938	230.169
					H4C	shoots		220.739	14.527	114.166	3.639	0.469	26.335
					H4	soil		6051.389	322.581	2200.248	107.556	5.301	2951.923
							<i>translocation factor</i>	20%	27%	17%	32%	24%	11%
							<i>accumulation factor</i>	11%	11%	18%	7%	23%	4%
H5	-10.8054	-76.2776	14.3	187.2	H5R	roots		610.061	139.950	603.379	17.745	1.891	189.553
					H5C	shoots		156.410	30.119	114.977	4.962	0.223	38.757
					H5	soil		4612.349	502.892	1829.869	157.731	11.024	1990.127
							<i>translocation factor</i>	26%	22%	19%	28%	12%	20%
							<i>accumulation factor</i>	8%	17%	20%	7%	10%	6%
B21	-11.1442	-76.0650	57.9	153.3	B21R	roots		9.806	11.068	27.210	0.501	17.157	1.815
					B21C	shoots		61.874	6.623	34.965	0.174	0.575	0.668
					B21	soil		788.770	15.838	50.895	2.728	0.247	151.769
							<i>translocation factor</i>	631%	60%	128%	35%	3%	37%
							<i>accumulation factor</i>	5%	56%	61%	12%	3593%	1%
B27	-11.1257	-76.1002	52.6	160.6	B27R	roots		2115.799	9.594	65.681	4.869	0.146	15.146
					B27C	shoots		346.357	3.423	18.126	0.909	0.054	1.624
					B27	soil		7023.413	8.154	66.623	24.514	0.084	71.412
							<i>translocation factor</i>	16%	36%	28%	19%	37%	11%
							<i>accumulation factor</i>	18%	80%	63%	12%	120%	12%
G21	-10.9784	-76.0337	41.6	143.4	G21R	roots		5455.532	17.430	47.971	13.527	0.916	18.526
					G21C	shoots		580.312	5.673	19.578	1.098	0.034	1.823
					G21	soil		5154.924	6.824	28.348	13.727	0.034	33.789
							<i>translocation factor</i>	11%	33%	41%	8%	4%	10%
							<i>accumulation factor</i>	59%	169%	119%	53%	1388%	30%
H22	-10.7687	-76.2687	10.1	182.0	H22R	roots		598.267	811.740	205.012	31.018	0.796	171.657
					H22C	shoots		359.598	490.727	267.503	19.694	0.307	114.060
					H22	soil		7210.474	2674.341	1476.765	233.901	4.601	2214.905
							<i>translocation factor</i>	60%	60%	130%	63%	39%	66%
							<i>accumulation factor</i>	7%	24%	16%	11%	12%	6%
G12	-10.8433	-76.2664	18.3	181.8	G12R	roots		1377.414	282.397	722.810	94.482	17.206	599.845
					G12C	shoots		332.305	67.459	277.916	22.501	7.466	149.528
					G12	soil		2827.369	623.739	1584.006	155.886	23.640	947.868
							<i>translocation factor</i>	24%	24%	38%	24%	43%	25%
							<i>accumulation factor</i>	30%	28%	32%	38%	52%	40%

*Table SI4:
U.S. EPA maximum contamination level for industrial waste and measured percentiles(P),
background concentration, and enrichment factors on the Junin Plain[^]*

	Pb	As	Cu	Zn	Ag
EPA maximum contamination level	420	75	4300	7500	N/A
median observed	133	28.3	23.6	85.7	0.235
max observed	21299	863	2674	17651	42.5
P90	3081	138	190	363	1.97
P80	758	82.9	86.5	194	0.831
P70	323	57.2	56.2	137	0.489
P60	200	38.3	36.6	114	0.359
P50	133	28.3	23.6	85.7	0.235
P40	107	20.7	17.2	67.1	0.154
P30	84.5	15.9	12.7	53.2	0.112
P20	57.5	11.0	8.88	41.3	0.070
P10	34.9	6.30	6.66	28.4	0.042
background: SP1-9*	23.0	1.76	19.9	5.16	0.020
enrichment factors: P90/SP1-9*	133.8	78.1	9.52	70.2	107

[^] all values are in mg/kg except for enrichment factors, which are dimensionless.

* background and resultant enrichment factor based on soil sample obtained 18 cm below the surface (Figure 2) at a site 4 km NNW of Cerro de Pasco (Figures 1 and 3).