

## Supplemental Materials

### Methods

#### *Sample locations, preparation and wet chemistry.*

The samples in this study were collected by deep submarine dredging from four seamounts (Papatua, Moki, Seamount D ["Dino"], Malulu) and one atoll (Rose) in the Samoan region. Several samples in this study were collected during the 2017 expedition (EX1702) aboard the NOAA *Okeanos Explorer*: Moki seamount (basaltic sample D7-2), Dino seamount (basaltic sample D11-1), Rose atoll (pillow fragment sample D3-2), and Malulu seamount (hyaloclastite sample D12-5). Finally, two samples from the 1999 AVON2/3 expedition aboard the R/V *Melville* which were previously characterized are presented here with modern Pb-isotopic analyses: AVON2/3-D67-11 from Malulu seamount which is an altered basalt, and basaltic sample AVON2/3-D66-1 from Rose atoll. Tables S1 and S2 present previously published data on these two Rose samples in addition to two other samples—AVON2/3-D65-18 (from Malulu seamount) and ALIA-DR129-05 (from Papatua seamount)—from non-Samoan seamounts in the Samoan region. Sample locations and geochemical characterization of these four previously published lavas can be found in **Jackson et al. (2010)**.

Samples were crushed in plastic bags to avoid exposure to metal. Crushed material was then sieved. For two samples (EX1702-D12-5 hyaloclastite from Malulu seamount and EX1702-D7-2 basalt from Moki seamount), clinopyroxene was removed for radiogenic isotopic work due to the lack of fresh basaltic material. For Rose atoll sample EX1702-D3-2, volcanic glass was removed from the pillow rim for radiogenic isotopic analysis (this glass was also characterized for major and trace element compositions, but we also present whole rock major and trace element data for this sample as well). For sample EX1702-11-1 (Seamount D), 200 mg of the freshest (0.5 to 1 mm) rock chips were analyzed, targeting groundmass. We also separated the freshest groundmass chips from two previously characterized lavas—AVON2/3-D66-1 (Rose atoll) and AVON2/3-D67-11 (Malulu seamount)—for a new characterization using modern Pb isotopic analyses and new analyses of Sr and Nd isotopes on the same material.

The groundmass samples (including the EX1702-D3-2 glass) were treated with a heavy leaching protocol described in **Price et al. (2016)** which, in addition to the 6N HCl leaching treatment, uses hot 4N HNO<sub>3</sub> and hot 30% H<sub>2</sub>O<sub>2</sub>. The clinopyroxene samples were first leached in concentrated HCl on a hot plate for 1 hour at 40° C, then in concentrated nitric for 1 hour at 40° C, with ~1 hour of sonication in the same acid following each leaching step; after this initial leach, the clinopyroxenes were subjected to the same "heavy leaching" as the groundmass and glass samples (and, following leaching, only pristine clinopyroxenes were selected for dissolution and analysis). Note that replicated analysis of the D11-1 groundmass followed additional leaching: it was treated with the strong leaching protocol followed by an additional 8 hours of leaching in 6N HCl at 60° C and an additional 10 minutes leaching in cold concentrated HF; this may explain the offset in Sr, Nd, and Pb isotopes between the original and replicate rounds of analyses of this sample. USGS reference materials were not leached. Following leaching, samples were rinsed and sonicated repeatedly in MilliQ H<sub>2</sub>O ( $\geq 18.2$  M $\Omega \cdot$  cm deionized water). Sample dissolution, wet chemistry (including Sr, Pb, Hf and Nd elemental separations), and mass spectrometry was carried out in one of the three following institutions:

1. One batch of samples underwent Sr, Nd, and Pb chemical separations at UCSB, with Sr and Nd isotopes analyzed on the UCSB TIMS and Pb isotopes analyzed on the WHOI

MC-ICP-MS; the wet chemistry follows methods developed in **Price et al. (2014)** with modifications as follows: Following dissolution in concentrated HF and HNO<sub>3</sub>, Sr and Pb were purified by two passes through 100 µL of Eichrom Sr resin (25-50 µm), and Nd purified was purified from the wash of the Sr resin using a two-step method employing Eichrom TRU resin (100-150 µm) followed by Eichrom LN-Spec resin (50-100 µm). Total procedural blanks are <200 pg for Sr, <50 pg for Nd, and < 120 pg for Pb.

2. A second batch of samples underwent Sr, Nd, and Pb chemical separations at UCSB as described above, with Sr and Nd isotopes analyzed on the UCSB TIMS. However, Pb isotopes were analyzed on the University of South Carolina MC-ICP-MS.
3. A third batch of samples, which included just the cpx separates for EX1702-D12-5 and EX1702-D7-5, underwent dissolution, chemical separations (for Sr, Nd, Hf, and Pb), and mass spectrometry at the University of South Carolina.

Further description of wet chemistry and mass spectrometry is provided below.

#### *Sr and Nd mass spectrometry at UCSB and Pb mass spectrometry at WHOI.*

Sr and Nd isotopes were analyzed on a Thermo Triton Plus TIMS mass spectrometer housed at UCSB. 500 ng of Sr or Nd was loaded on outgassed, zone-refined Re (99.999% purity, H-Cross, USA) filaments. With the exception of the first <sup>87</sup>Sr/<sup>86</sup>Sr analyses (EX1702-D12-5 clinopyroxene, EX1702-D7-5 clinopyroxene, EX1702-D11-1, EX1702-D3-2) and associated standards, which used a 33 picoamp gainboard but did not employ amplifier rotation, all other Sr and Nd isotopic analyses of samples, replicates, and associated standards employed amplifier rotation on 10<sup>11</sup> ohm amplifiers and a 3.3 picoamp gainboard. Gains were run with the start of a new barrel. Approximately 20% of analysis time was devoted to baselines: baselines are taken with each rotation of the amplifiers (i.e., because 5 amplifiers-cup pairs were used during analyses, 5 baselines were taken during each full amplifier rotation). Intensities were kept at approximately 3V on mass 88 and 3V on mass 144 during <sup>87</sup>Sr/<sup>86</sup>Sr and <sup>143</sup>Nd/<sup>144</sup>Nd analyses, respectively. Sr and Nd isotopes were corrected for mass bias assuming an exponential law and using canonical <sup>86</sup>Sr/<sup>88</sup>Sr and <sup>146</sup>Nd/<sup>144</sup>Nd ratios of 0.1194 and 0.7219, respectively. Isobaric interferences from Rb and Sm were corrected by monitoring masses 85 and 147, but corrections to the <sup>87</sup>Sr/<sup>86</sup>Sr and <sup>143</sup>Nd/<sup>144</sup>Nd ratios were nominal. USGS reference materials (processed through all steps of wet chemistry and column chemistry with unknowns at UCSB) and sample unknowns were corrected for the offset between preferred and measured standard (NBS987 or JNdi) values with each barrel: preferred value for NBS987 <sup>87</sup>Sr/<sup>86</sup>Sr is 0.710240, and JNdi is 0.512099 (**Garçon et al., 2018**). On the UCSB Triton Plus the average <sup>87</sup>Sr/<sup>86</sup>Sr and <sup>143</sup>Nd/<sup>144</sup>Nd and long-term reproducibility, up to and including this study, of NBS987 and JNdi using amplifier rotation is 0.710246 ± 0.000011 (2SD, N=29) and, 0.512100 ± 0.000004 (2SD, N=27), respectively. The corresponding average <sup>87</sup>Sr/<sup>86</sup>Sr and long-term reproducibility when analyzing NBS987 without amplifier rotation is 0.710244 ± 0.000014 (2SD, N=39).

We note that prior analyses of the AVON2/3 cruise samples from Rose (AVON2/3-D66-1) and Malulu (AVON2/3-D67-11), reported in **Jackson et al. (2010)**, should be replaced by new analyses of these two samples shown in **Table S1**, for two reasons: 1) the previously published Pb isotopic analyses were made by TIMS without a spike addition to control for in-run mass fractionation and 2) these two samples exhibit significant alteration and acid leaching for the prior analyses may not have been sufficient to have removed alteration phases. We note that the new <sup>143</sup>Nd/<sup>144</sup>Nd analysis for AVON2/3-D67-11 (0.512982) shows significant disagreement with the prior analysis (0.512796, after correction to the JNdi reference frame use here by

applying the La Jolla to JNdi conversion from **Tanaka et al., 2000**) made at WHOI in the year 2000 and reported by **Jackson et al. (2010)**. In order to evaluate the accuracy of the new  $^{143}\text{Nd}/^{144}\text{Nd}$  measurement, three additional aliquots of this sample were obtained from the WHOI dredge repository, including the original bag of crushed rock chips for this sample from which material was extracted for the 2000 analysis (see AVON2/3-D67-11 rep1, rep2, and rep3 in **Table S1**); new batches of chips were prepared by separately crushing each aliquot, and the different aliquots of chips were leached and underwent wet chemistry and mass spectrometry during a separate session than the original analyses. The three replicate  $^{143}\text{Nd}/^{144}\text{Nd}$  analyses (0.512982, 0.512980, and 0.512981) show excellent agreement with the new  $^{143}\text{Nd}/^{144}\text{Nd}$  result, confirming its accuracy. We note that the new analyses of Sr and Pb isotopes for this sample are similar to the published data for this sample reported in **Jackson et al. (2010)**.

Using the Pb fractions purified at UCSB, Pb isotopic analyses were carried out at the Woods Hole Oceanographic Institution using the Thermo Neptune MC-ICP-MS housed there (**Hart and Blusztajn, 2006**). Fractionation correction was made by Tl-addition assuming an exponential fractionation law (**White et al., 2001**). Samples and an aliquot of AGV-2 (processed through all steps of column chemistry and mass spectrometry with the samples) were corrected for the offset between preferred (i.e., values from **Eisele et al., 2003**; ( $^{206}\text{Pb}/^{204}\text{Pb} = 16.9409$ ,  $^{207}\text{Pb}/^{204}\text{Pb} = 15.4976$ , and  $^{208}\text{Pb}/^{204}\text{Pb} = 36.7262$ )) and measured ratios of NBS981.

For a different subset of samples (specified in **Table S2**), Pb fractions purified at UCSB were carried out at the University of South Carolina on Thermo Neptune MC-ICP-MS housed there. The samples, together with an aliquot of BCR-2 (processed through all steps of column chemistry and mass spectrometry with the samples) were corrected for the offset between preferred (i.e., values from **Eisele et al., 2003**) and measured ratios of NBS981.

*Sr, Nd, Hf and Pb isotopic and major and trace element concentrations on clinopyroxenes from samples EX1702-D12-5 and EX1702-D7-5 at U. South Carolina.*

Following leaching, visually fresh clinopyroxenes were picked under a binocular microscope. The clinopyroxene samples were then processed at the Center for Elemental Mass Spectrometry, University of South Carolina. Approximately 150-200 mg of sample was first leached (using the same protocol described above) and dissolved in Teflon distilled HF:HNO<sub>3</sub> (3:1) mixture on a hot plate for ~ 3 days, with frequent sonication. After drydown the samples were picked in 6N HCl with added boric acid in 10N HCl to complex fluorides, which increases yields in Hf chemistry (**Frisby et al., 2016**). Afterwards the samples were converted to nitrates. A small aliquot (~5%) was taken for Sr and Nd isotopes and the remainder was dried down with HBr for Pb and then Hf chemistry. A precisely determined aliquot of the dissolved clinopyroxene samples was pipetted from the solution and gravimetrically diluted for trace element analyses prior to the splitting of Sr-Nd and Pb-Hf fractions. Trace elements were then analyzed by ICP-MS with an aliquot of BHVO-1 on the Element2 following established methods for the lab (e.g. **Frisby et al, 2016**).

For the chemical separations, Sr was separated first on an Eichrom Sr-spec resin and the washes containing the rest of the elements were processed through an Eichrom TRU spec resin to concentrate the LREEs, and then on an Eichrom Ln-Resin to isolate Nd (**Frisby et al, 2016**). The Pb was separated on anion resin in HBr and HCl media, and the washes from that were processed for Hf following the method of **Munker et al. (2001)**. An unleached BCR-2 powder was dissolved and processed through all steps of chemistry and mass spectrometry with sample unknowns. Total procedural blanks during the analytical session were 10 pg for Pb, 180 pg for

Sr, <10 pg for Nd, and 40 pg for Hf.

The isotopic compositions of Sr, Nd, Pb, and Hf were measured on the Thermo Neptune housed at the University of South Carolina following methods provided in **Beguelin et al., (2017)**. All samples were corrected for mass bias using the exponential law: Pb was corrected using the Tl addition method (**White et al., 2001**),  $^{87}\text{Sr}/^{86}\text{Sr}$  was corrected using  $^{86}\text{Sr}/^{88}\text{Sr}$  of 0.1194,  $^{143}\text{Nd}/^{144}\text{Nd}$  was corrected using  $^{146}\text{Nd}/^{144}\text{Nd}$  of 0.7219, and  $^{176}\text{Hf}/^{177}\text{Hf}$  was corrected using  $^{179}\text{Hf}/^{177}\text{Hf}$  of 0.7325. All isotopic data on samples were corrected for the offset between measured and preferred standards using the following preferred values: SRM981 values from **Eisele et al. (2003)**; NBS987  $^{87}\text{Sr}/^{86}\text{Sr}$  value of 0.710240; JMC-475  $^{176}\text{Hf}/^{177}\text{Hf}$  value of 0.282160; JNdi  $^{143}\text{Nd}/^{144}\text{Nd}$  value of 0.512099 (**Garçon et al., 2018**).

During the course of these measurements, reproducibility of  $^{87}\text{Sr}/^{86}\text{Sr}$  on NBS987 was 22 ppm, of  $^{143}\text{Nd}/^{144}\text{Nd}$  on JNdi-1 was 20 ppm, of  $^{176}\text{Hf}/^{177}\text{Hf}$  on JMC-475 was 28 ppm, and Pb isotopic compositions on NBS981 were 62ppm on  $^{206}\text{Pb}/^{204}\text{Pb}$ , 68 ppm for  $^{207}\text{Pb}/^{204}\text{Pb}$ , and 80 ppm for  $^{208}\text{Pb}/^{204}\text{Pb}$  (all 2SE). An unleached BCR-2 powder was run together with the clinopyroxenes through all steps of column chemistry and mass spectrometry, and data are shown in **Table S1**.

#### *Whole rock major and trace element analyses at Washington State University.*

For samples EX1702-D7-2, EX1702-D3-2, and EX1702-D11-1, 10 to 20 g blocks of rock were cut with the rock saw, and care was taken to avoid visibly altered portions of rock. The blocks were cleaned with silicon carbide sand paper, and sonicated in MilliQ H<sub>2</sub>O. Samples were crushed and then powdered in an agate shatterbox (with cleaning with silica between barrels) at the Geoanalytical lab at the Washington State University (WSU). Major and trace element concentrations were obtained at WSU by X-ray fluorescence (XRF) and ICP-MS. XRF and ICP-MS methods, and evaluation of accuracy using international standards, are reported elsewhere (**Knaack et al., 1994; Johnson et al., 1999**), but summarized briefly here. The precision (1 $\sigma$ ) for major elements in basalts by XRF is 0.11–0.33% (1 $\sigma$ ) of the amount present for SiO<sub>2</sub>, Al<sub>2</sub>O<sub>3</sub>, TiO<sub>2</sub>, P<sub>2</sub>O<sub>5</sub>) and 0.38–0.71% for other elements. Trace element precision of basalts by ICP-MS is 0.77–3.2% (1 $\sigma$ ) for trace elements except for U (9.3%) and Th (9.5%). An aliquot of the USGS reference material BCR-2 was analyzed as an unknown together with the basaltic unknowns, and the data are reported in **Table S2**. The new analysis of BCR-2 reported here is compared to major and trace element compositions for this reference material reported in **Jochum et al. (2016)**. Whole rock major and trace element analyses of the AVON2/3-D67-1, AVON2/3-D66-1, AVON2/3-D65-18 and ALIA-DR129-05 were made following the same methods and data are reported in **Jackson et al. (2010)**.

#### *In situ major element analyses on glass by electron microprobe at UC Santa Barbara.*

Glass was available for EX1702-D3-2. Sample chips were analyzed by electron microprobe at UC Santa Barbara using primary standards and following analytical conditions outlined in **Jackson et al. (2015)**. The MORB basaltic secondary standard 519-4-1 was analyzed repeatedly throughout the analytical session. Major element compositions for sample unknown glass and the secondary standard (and previously published data on this secondary standard from **Melson et al., 2002**) are reported in **Table S2**.

*In situ trace element analyses by LA-ICP-MS at Clermont-Ferrand.*

Trace element analyses on the EX1702-D3-2 glass sample were made using a laser ablation ICP-MS system housed at Laboratoire Magmas et Volcans at Clermont-Ferrand. Analytical methods are outlined in **Oulton et al. (2016)** and **Reinhart et al. (2018)**. Analyses were made using a Thermo Scientific Element XR ICP-MS coupled to a Resonetics M-50E 193 nm ArF excimer laser.  $^{43}\text{Ca}$  was used as an internal standard. All surfaces were preablated (for 1 s at 10 Hz) prior to analysis. Analyses of samples and standards used a 47  $\mu\text{m}$  laser spot, and the laser was fired with a 4 Hz repetition rate. Analyses were conducted over 80 second ablation periods, with 20 seconds of blank analysis (with the laser off) before samples analysis and (following a washout period) 20 seconds of blank analysis (again, with the laser off) after the analysis. Analyses were made in low resolution mode using triple mode with a 20% mass window and a 20 ms integration window. Acceleration voltage was scanned between magnet scans to ensure peak positions were maintained. Calibration curves were generated using NIST612 (**Gagnon et al., 2008**) and BCR-2 (**Jochum et al., 2006**) glass. Replicate analyses of a MORB glass, 519-4-1, were made throughout the analytical session to monitor precision and accuracy of analyses. The reproducibility of the trace element analyses was better than 10% (2RSD, N=8) for all elements except for Cs (44%). Measured concentrations are compared with previously published analyses from **Gale et al. (2013)** in **Table S2**.

*$^{40}\text{Ar}/^{39}\text{Ar}$  Geochronology at Oregon State University*

Clinopyroxene separates were prepared from basalt samples AVON-D66-1 (Rose atoll) and EX1702-D7-2 (Moki) following the methods outlined in Konrad et al. (2019). Separate gases were processed and analyzed on a new extraction line and ARGUS VI mass spectrometer housed at Oregon State University. Incremental heating experiments consisted of 20–21 steps using a  $\text{CO}_2$  laser. Blanks were analyzed at the experiment start, end and between every three heating steps. Gas was processed and analyzed using the same methods outlined for the ARGUS-VI-E in Konrad et al. (2019). ArArCalc v.2.7.0 (Koppers, 2002) was used to process results and calculate age determinations. A Fish Canyon Tuff fluence monitor age of  $28.201 \pm 0.046$  Ma ( $2\sigma$ ; Kuiper et al., 2008) and a total decay constant of  $5.530 \pm 0.097 \times 10^{-10}$  year $^{-1}$  ( $2\sigma$ ; Min et al., 2000) were used to calculate the ages.

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## Figure Captions

**Figure S1. Isotopic data for volcanoes in this study plotted with existing data for Macdonald and Arago volcanoes.** The Moki and Malulu isotopic data produced for this study are from clinopyroxene phenocrysts, as seawater alteration prevented the analysis of whole rock samples. New data also include pillow rim glass analyses from Rose atoll, whole rock data from Dino seamount, and new whole rock data on Rose and Malulu lavas. Whole rock data for the Rose atoll and the Malulu and Papatua seamounts was published by Jackson et al. (2010). Young (Cook-Austral) Arago and Macdonald isotope fields are represented by red and blue backgrounds, respectively, and solid outlines. Old (Cretaceous) Arago and Macdonald fields are represented by light and dark gray fields with long and short dashed outlines, respectively.  $^{143}\text{Nd}/^{144}\text{Nd}$  data from sample SUA-1 (Konter et al., 2008) is not shown due to relatively large measurement errors ( $\pm 195$  ppm). The Tuvalu, Gilbert, Marshall and Wake islands and seamounts define the older portion of the Arago hotspot (data sources from Staudigel et al., 1991; Konter et al., 2008; Finlayson et al., 2018; Konrad et al., 2018); the Tokelau Islands and seamounts define the older portion of the Macdonald hotspot (data sources from Konter et al., 2008). Measured isotopic values are shown, but where  $^{40}\text{Ar}/^{39}\text{Ar}$  ages are available (Moki, Rose atoll), an age correction to the time of eruption is calculated. The isotopic change due to the age correction is shown by the line extending from the respective datapoints (which represents the magnitude and direction of the age correction). The other samples (Malulu, Papatua, and Dino) are not associated with ages and age corrections are not provided. All data not produced by this study was previously published and downloaded from Georoc (<http://georoc.mpch681mainz.gwdg.de/georoc>). Figure modified after Jackson et al. (2020).

**Figure S2. Trace element spider diagrams for whole rock and clinopyroxene samples from study volcanoes.** Trace element concentrations are normalized to pyrolite from McDonough and Sun (1995). Whole rocks are shown in the top panel, and clinopyroxenes in the lower panel.

**Figure S3:** Age plateaus and inverse isochrons for clinopyroxene (CPX) separates from AVON-66-1 (Rose Atoll; left panel) and EX1702-D7-2 (Moki Seamount; right panel). The Rose Atoll sample is a combined incremental heating result for a 355-500  $\mu\text{m}$  size fraction (blue) and a 500-595  $\mu\text{m}$  size fraction (orange), which is justified because both plateau ages are concordant; the plateau age is adopted for the Rose age determination. A single heating experiment is presented for the Moki Seamount clinopyroxene (cpx) that contained very low concentrations of K and corresponding radiogenic gas, resulting in high individual step uncertainties, particularly for the first two heating steps. The lines above the plateaus represent steps used in the age calculation, and all steps were incorporated into the respective ages. Inverse isochrons are shown beneath the plateau panels for each analysis. Black squares represent points used in the calculations while white squares represent excluded points. The inverse isochron age determination is preferred (for Moki) due to the slightly above atmospheric  $^{40}\text{Ar}/^{36}\text{Ar}_{\text{int}}$  value. Both measurements meet standard  $^{40}\text{Ar}/^{39}\text{Ar}$  quality criteria with plateaus consisting of  $>60\%$  cumulative  $^{39}\text{Ar}_{(\text{K})}$ , appropriate mean-square of weighted deviants (MSWD) and probability-of-fit (P) values greater than 0.05 (see Supplemental Tables 3 and 4).

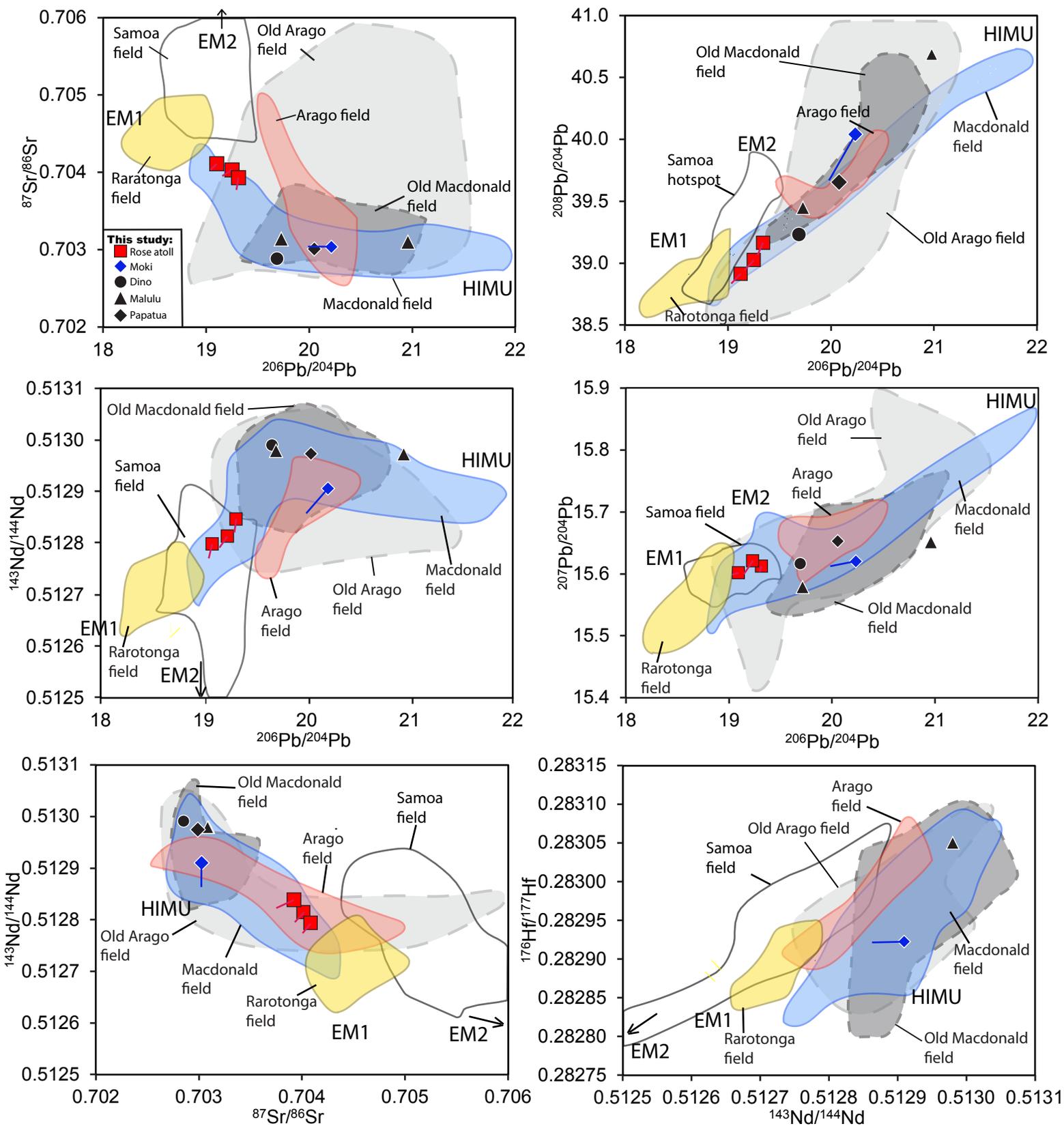


Figure S1

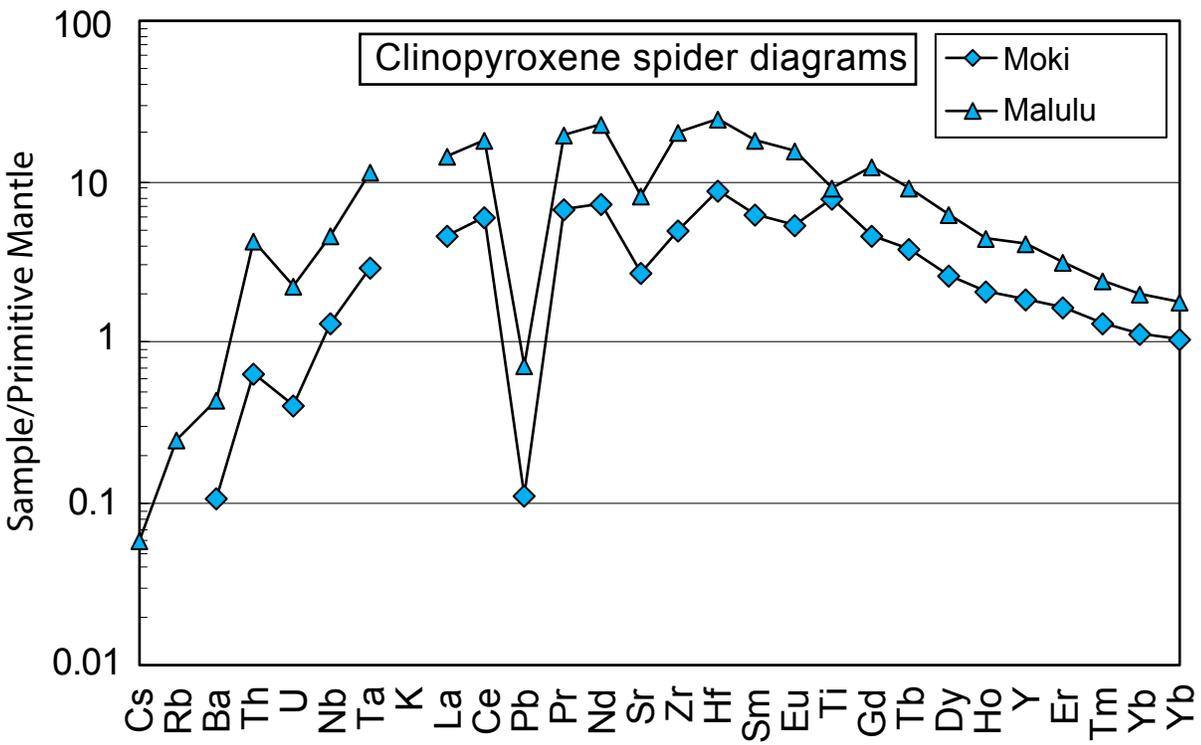
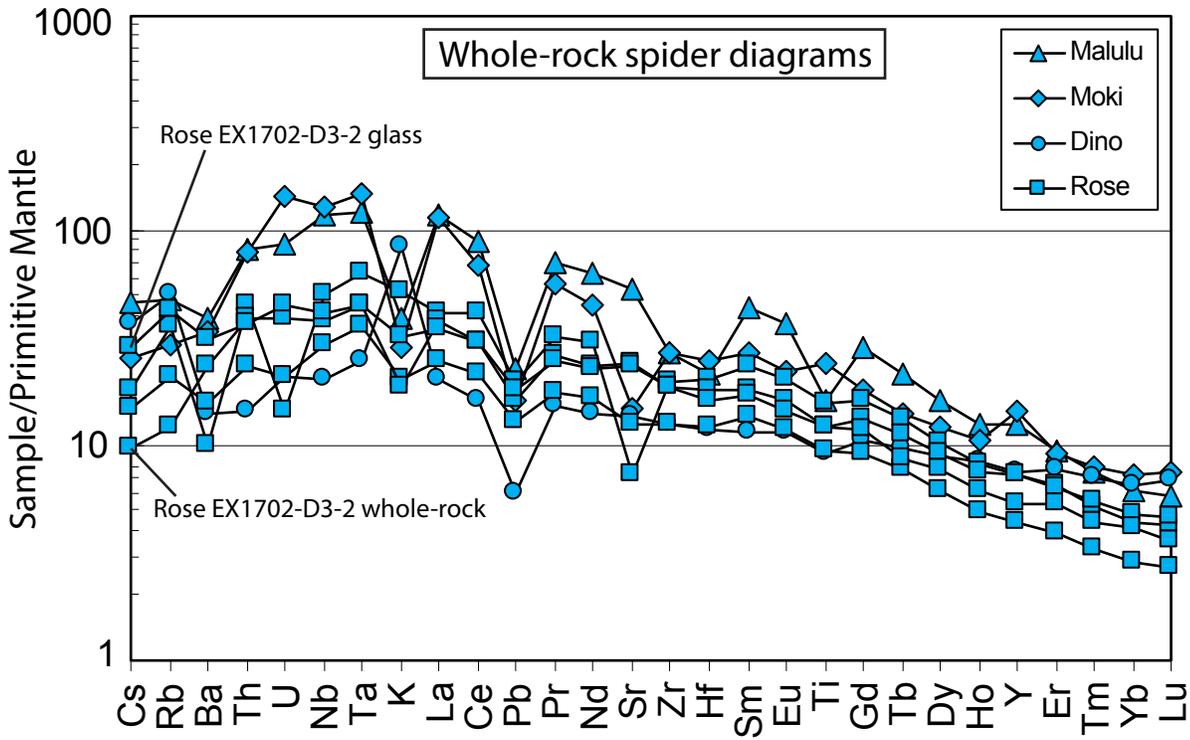


Figure S2

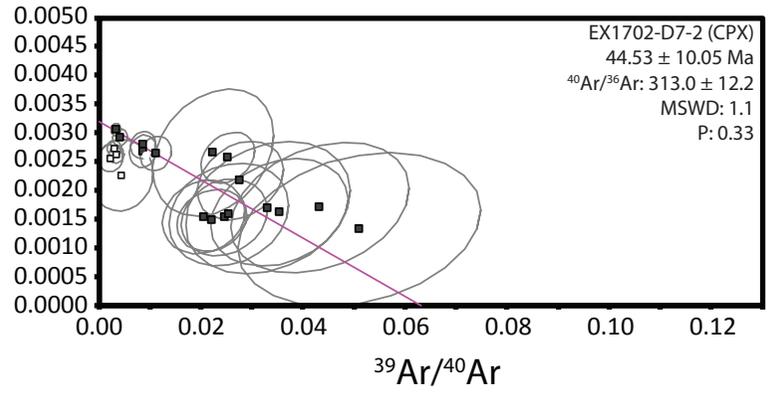
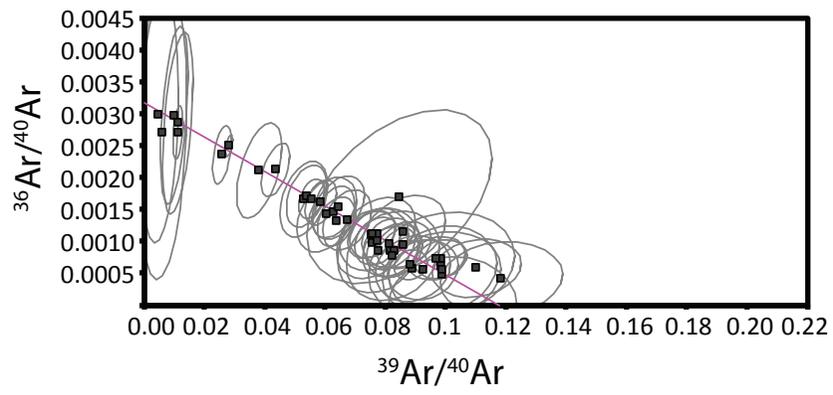
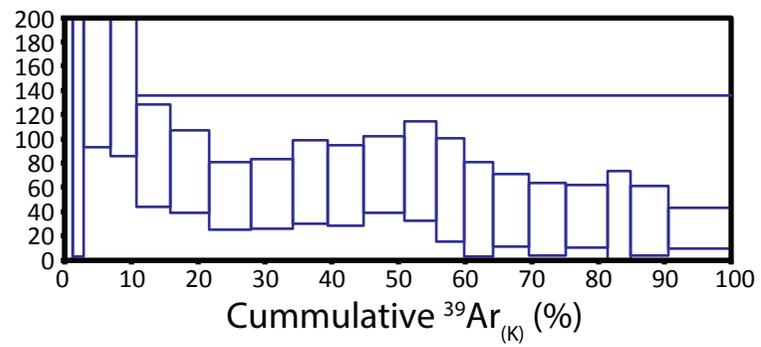
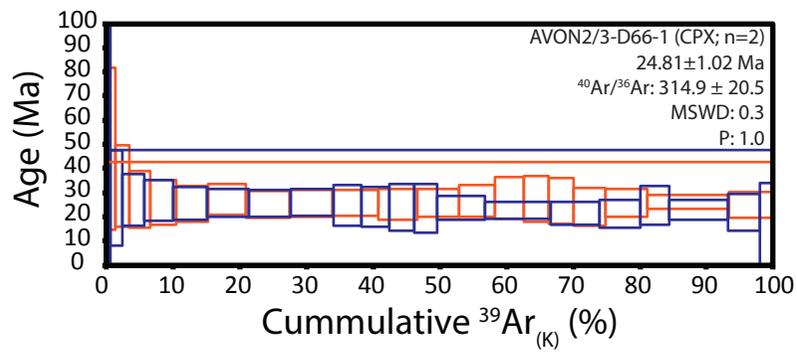


Figure S3